

Original article

Evaluation of Long Term Intake of *Citrus aurantifolia* Juice on Fertility Indices in Male Wistar Rats

Zainab Ahmad¹, Hauwa Abdullahi^{1*} , Natasha Adamou¹, Hauwa Umar², Sharif Alhassan³

¹Department of Obstetrics and Gynecology, Faculty of Clinical Sciences, Bayero University Kano, Nigeria

²Department of Obstetrics and Gynecology, Ahmadu Bello University Zaria, Kaduna, Nigeria

³Department of Medical Microbiology and Parasitology, Faculty of Clinical Sciences, Bayero University Kano, Nigeria

ARTICLE INFO

Corresponding Email. drhauwa@yahoo.com

Received: 12-01-2023

Accepted: 10-02-2023

Published: 17-02-2023

Keywords. Infertility, *Citrus Aurantifolia*, Hormones, Spermatogenesis, Wistar Rats.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>

ABSTRACT

Background and aims. Infertility is one of the most common presentations in gynecological clinics. Male-related factors account for over 30% of all cases. Several factors are known to interfere with fertility promoters and indices, such as drug treatment, environmental toxins, air pollution and stress. *Citrus aurantifolia* was shown to affect the number of shed ova and sperm cells. This study aimed to determine the effects of long-term intake of *Citrus aurantifolia* juice (CAJ) on sperm parameters and sex hormones in male Wistar rats. **Methods.** A total of twenty-four (24) rats were used in the study and were randomly subdivided into 4 groups (n=6). Group I (normal control) received 1 ml/kg distilled water. Group II received 1 ml/kg concentrated CAJ (cCAJ), Group III received 1 ml/kg diluted CAJ (dCAJ) at a 1:1 ratio, and Group IV received 1 ml/kg dCAJ at a 1:2 ratio. The preparations were given daily by gavage for a period of 42 days. Blood and semen samples collected from each rat were subjected to hormonal and seminal fluid analysis. **Results.** The hormonal assay revealed a significant decrease in prolactin (p=0.0014) and LH (p=0.0124) levels with a significant increase in FSH (p=0.0021) and testosterone (p=0.001) levels in Group II that received cCAJ compared with Group I. The seminal fluid analysis revealed that the mean values of active cells, viable cells, total counts and normal cells in Group II were significantly lower (p value=0.0001) than those in the normal group. However, the mean values significantly increased (p value=0.001) in Groups III and IV but were still lower than those in Group I. **Conclusion.** The findings of this study indicate a negative effect of the CAJ on male fertility indices with a possible effect on spermatogenesis that may lead to infertility in male adult Wistar rats.

Cite this article. Ahmad Z, Abdullahi H, Adamou N, Umar H, Sharif A. Evaluation of Long Term Intake of *Citrus Aurantifolia* Juice on Fertility Indices in Male Wistar Rats. *Alq J Med App Sci.* 2023;6(1):37-43. <https://doi.org/10.5281/zenodo.7650057>

INTRODUCTION

Infertility is a major concern for individuals, families and health care givers in many societies throughout the world. Male-related factors account for over 30% of all cases, and several factors are known to interfere with fertility promoters and indices, such as drug treatment, environmental toxins, air pollution and stress [1]. *Citrus aurantifolia*, which is popularly known as lime, belongs to the genus of flowering plants in the family Rutaceae and is a common edible fruit of this genus. The entire lime plant has been demonstrated to have a wide range of uses, including medicinal, industrial, cosmetic, and pharmaceutical uses. The *Citrus aurantifolia* fruit is typically round, lime green in color, 3–6 centimeters (1.2–2.4 in) in diameter, and contains acidic juice vesicles [2].

The bioactive constituents of *Citrus aurantifolia* include flavonoids, limonoids, alkaloids, ascorbic acid, and tannins [3]. *Citrus aurantifolia* is readily available in every part of our society with its increasing social and medicinal uses by both men and women. In many communities, women use CAJ as a barrier contraceptive, and there is a reported history of African women douching with *Citrus aurantifolia* juice (CAJ), lemon juice, vinegar or acidic soft drinks on the belief that it may prevent pregnancy and/or sexually transmitted diseases [4].

Reproduction in males is a complex process that is coordinated and controlled by reproductive hormones such as testosterone, luteinizing hormone and follicle-stimulating hormone. The anterior pituitary controls reproductive processes through the secretion of gonadotropins such as luteinizing hormone (LH) and follicle stimulating hormone (FSH). Gonadotropins indirectly stimulate endogenous testosterone production and directly control spermatogenesis by the testes [5]. In men, high serum prolactin concentrations decrease gonadotropin secretion, thus decreasing testicular function and resulting in low serum testosterone concentrations. The major symptoms are loss of sexual desire, erectile dysfunction, muscle weakness, and infertility. Highly elevated levels of prolactin decrease the levels of sex hormones, i.e., estrogen in women and testosterone in men [6]. Prolactin acts in a cytokine-like manner and acts as an important regulator of the immune system. Prolactin has important cell cycle-related functions as a growth differentiating and anti-apoptotic factor [7]. As a growth factor binding to cytokine-like receptors, it also has a profound influence on hematopoiesis and angiogenesis and is involved in the regulation of blood clotting through several pathways [8]. Testosterone is very useful in the completion of the spermatogenic process, as it enhances the conversion of round spermatids to spermatozoa during spermatogenesis [9,10]. This study, therefore, assessed the effect of the long-term intake of juice on the fertility indices of male Wistar rats.

METHODS

Extract preparation

One hundred fresh fruits of *Citrus aurantifolia* were obtained from the Rimi Market in Kano, Kano State, Nigeria. The fruits were properly washed and sliced into two halves each. The juice was extracted using a juice extractor and filtered through a sieve, and the residual pulp and seeds were discarded. The CAJ extracts were pooled and collected into small clean plastic bottles, covered and refrigerated at 4°C for experimental use.

Ethical considerations

Ethical approval for the study was obtained from the ethical committee of Bayero University Kano (College of Health Sciences). Following approval, the researchers ensured that laboratory animals were treated according to high ethical and scientific standards.

Experimental animals

Twenty-four (24) adult male Wistar rats weighing between 110 and 130 g were procured from the Animal House of the Department of Pharmacology and Therapeutics of Ahmadu Bello University (ABU) Zaria. Principles of Laboratory Animal Care (NIH Publication No. 85-23, Revised 1985) were followed, as well as specific national laws where applicable. All experimental methods were examined and approved by the ethical committee of the College of Health Sciences, Bayero University, Kano. The Wistar rats were kept at the animal house facility of the Department of Pharmacology, Bayero University Kano, for one week to acclimatize. The wister rats were fed standard diets (Vital Feeds and Grand Cereals obtainable from Brand Cereals and Oil Mills Ltd, Bukuru, Jos, Nigeria), and water was given ad libitum and maintained under standard conditions.

Experimental procedure

Twenty-four (24) rats were used in this experiment and were randomly subdivided into 4 groups (n=6). The wister rats in Group I (normal control) received 1 ml/kg distilled water only. Group II received 1 ml/kg body weight cCAJ, Group III received 1 ml/kg dCAJ at a 1:1 dilution, and Group IV received 1 ml/kg dCAJ at a dilution of 1:2. The preparations were given by gavage orogastrically daily using a metal cannula at 0900 hours for a period of 42 days.

Animal sacrifice, sample collection and analysis

Animals were sacrificed after chloroform anesthesia 24 hours after the last administration of cCAJ, dCAJ or distilled water. Blood samples from each animal were collected immediately through the jugular veins of each rat and transferred to serum separating tubes for hormonal assays. The SST bottles were properly labeled to allow easy identification. The blood samples were then conveyed to the chemical pathology laboratory for analysis using Wondfo Finecare Fia meter plus (model no. FS-113). The sperm were released by cutting the cauda epididymides longitudinally with a pair of fine-pointed scissors and compressing with forceps, and the sperm were therefore deposited free of epididymal tissue into the cavity. The sperm suspension was drawn into a plain container and conveyed to the laboratory within 10 to 15 minutes of collection for analysis using a Sperm Quality Analyzer (SQA IIIC-P, USA).

Statistical analysis

All results are presented as the mean \pm standard error of the mean. Three sets of data were analyzed using one-way ANOVA, followed by the least significant difference (LSD) procedure for significant F values at $P < 0.05$ using Excel and Statistical Package for Social Sciences (SPSS) version 22.

RESULTS

Effects of long-term administration of different concentrations of CAJ on gonadotrophic hormones

Forty-two days after oral administration of water for the control group (Group I) and fresh extract of cCAJ for Group II and 1:1 vs 1:2 dCAJ for Group III and Group IV, respectively (Table 1). The mean value of prolactin decreased significantly ($p=0.0014$) in Group II and Group III (0.013) compared with Group I. However, the prolactin level was not significantly different ($p=0.1855$) in Group III and Group IV compared with Group II. However, the testosterone level significantly increased ($p=0.001$) in Group II compared with Group I, and there was also a significant difference ($p=0.017$) in the mean values of testosterone in Group III and Group IV compared with Group II. The mean value of FSH among the Wistar rats in Group II that received cCAJ at a dose of 1 ml/kg was significantly ($p=0.0021$) higher when compared with Group I. There was also a significant decrease ($p=0.0223$) in the FSH level among Group III that were fed dCAJ at a 1:1 ratio and among Group IV ($p=0.0017$) that received 1:2 dCAJ when compared to Group II. The mean values of LH decreased significantly ($p=0.0124$) in Group II when compared with Group I. Likewise, the decrease in LH level in Group III and Group IV was significantly different ($p=0.0107$) from Group II.

Table 1. Effect of long-term oral administration of CAJ at different concentrations on sex hormones

Groups	Doses 1 ml/kg	Prolactin (μ /L)	Testosterone (ng/ml)	FSH (μ /L)	LH(μ /L)
Group I	Water	3.86 \pm 0.44	1.29 \pm 0.04	1.09 \pm 0.12	1.96 \pm 0.11
Group II	cCAJ	2.13 \pm 0.04 [#]	2.88 \pm 0.06 [#]	1.71 \pm 0.13 [#]	1.54 \pm 0.08 [#]
Group III	dCAJ (1:1)	2.55 \pm 0.39 [#]	1.83 \pm 0.07 ^{##}	1.34 \pm 0.13 ^{##}	1.60 \pm 0.06 ^{##}
Group IV	dCAJ (1:2)	2.86 \pm 0.47	1.39 \pm 0.05 ^{##}	1.28 \pm 0.42 ^{##}	1.75 \pm 0.06 ^{##}

LH=Luteinizing hormone, FSH= Follicle stimulating hormone, cCAJ= concentrated Citrus aurantifolia, dCAJ= diluted Citrus aurantifolia. The results are expressed as the mean \pm SE, $n = 6$, Values with superscript (#) are significantly different from the control (Group I). Values with superscript (##) are significantly different from Group II that received cCAJ.

Effect of long-term administration of different concentrations of CAJ on sperm parameters of Wistar rats

The effects of the CAJ in wister rats on semen parameters, such as active cell, sluggish cells, nonmotile cells, viable cell, nonviable cells, total counts, tail defects, head defects and normal cell were determined in this study.

In Figure 1, the total count decreased significantly (A, p -value=0.018) in Group II that received cCAJ when compared with Group I; likewise, the mean value of viable cells was also observed to have decreased in Group II and was significantly different (B, p -value= 0.032) from Group I. The non-viable cells were also observed to have increased significantly in Group II and Group III when compared with Group I (C, p value= 0.0001). There was also a significant difference (C, p -value= 0.001) in the mean values of non-viable cells in Group III and Group IV when compared with Group II. The active cells were also observed to have decreased significantly (D, p -value=0.0001) in Group II and Group III when compared with Group I. It was also observed that the active cells in Group IV was significantly different (D, p value=0.001) from that in Group II. The mean value of sluggish cells was observed to have increased in Group II and was significantly different (E, p - value=0.023) when compared with Group I. The non-motile cells increased significantly (F, p - value= 0.0001) in Group II and Group III when compared with Group I. It was also observed that the mean values of non-motile cells in Group III and Group IV were also significantly different (F, p -value=0.0001) from Group II.

Figure II displays the effect of the CAJ on sperm cell morphology. The sperm cells with tail defect were observed to have increased significantly (A, p -value=0.0001) in Group II when compared with Group I. Moreover, the mean values of sperm cells with tail defect in Group III and Group IV were observed to be significantly different (A, p -value= 0.001) from Group II. Sperm cells with head defect were observed to increase significantly (B, p -value=0.0001) in Group II when compared to Group I. The mean values of sperm cells with head defect were also observed to significantly differ between Group III and IV from Group II (B, p -value=0.006) and between Group III and Group IV (B, p -value=0.002). The mean values of sperm cells with normal cell morphology were observed to have decreased significantly (C, p -

value= 0.0001) in Group II and Group III when compared with Group I. There were also significant changes (C, p-value= 0.001) in the mean values of normal cells in Group IV when compared with Group II and Group III.

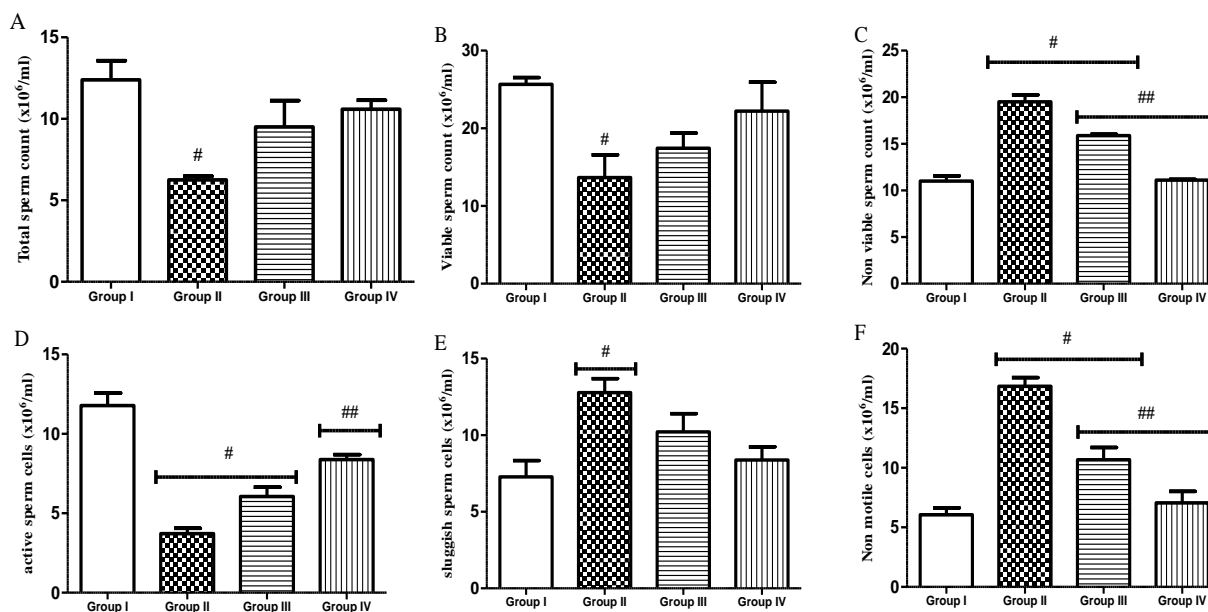


Figure 1. Effect of long-term intake of CAJ on sperm parameters. (A) Total sperm count, (B) viable sperm count, (C) nonviable sperm count, (D) active sperm cells, (E) sluggish sperm cells, and (F) non-motile cells. Bars with # are significantly different from Group I (P value = 0.018(A), 0.032 (B), 0.0001(C, D, F), 0.023 (E) and ## are significantly different from Group II (P values = 0.0001 (C, D, F).

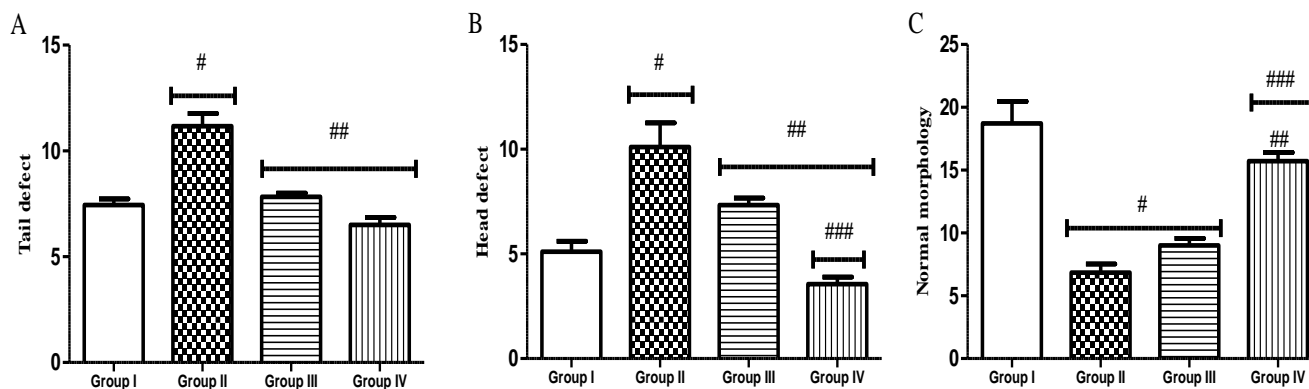


Figure 2. Effect of long-term intake of CAJ on sperm morphology. (A) tail defect (B) head defect (C) normal morphology. Bars with # differ significantly (p values = 0.0001 (A, C), and 0.006 (B)) from Group I, ## differ significantly (p values 0.001 (A, B, C)) from Group II and ### differ significantly (p value= 0.002 (B) and 0.001 (C)) from Group III.

DISCUSSION

Follicle stimulating hormone (FSH) is known to regulate the development, growth, pubertal maturation, and reproductive processes of the body. It stimulates primary spermatocytes to undergo the first division of meiosis to form secondary spermatocytes. It enhances the production of androgen-binding protein by the Sertoli cells of the testes by binding to FSH receptors on their basolateral membranes [11], and it is critical for the initiation of spermatogenesis. When the seminiferous tubules fail to produce sperm, secretion of FSH by the anterior pituitary increases. Conversely, when spermatogenesis proceeds too rapidly, pituitary secretion of FSH diminishes, which occurs as a result of negative feedback on the anterior pituitary by inhibin [12]. There was a significantly elevated FSH level in the group administered fresh CAJ (Group II). An increase in FSH has been reported to occur in patients with severely impaired spermatogenesis [13]. Therefore, CAJ can be said to be a weak anti-androgen that could have completely prevented FSH from binding

to its receptor, and as a result, the action of Sertoli cells is inhibited, which in turn adversely affects spermatogenesis and thus leads to elevated serum FSH by a negative feedback response in Group II. However, this finding seems to contradict reports by [14], which showed no significant changes in the FSH levels of male Wistar rats when administered CAJ, and [15], where there was a significant decrease in FSH levels in female rats that in turn reduced follicular development and ovulation upon the administration of CAJ. This indicates that the effect of CAJ on FSH levels could be sex dependent.

The primary role of LH in males is to stimulate the production of testosterone by Leydig cells. The mechanism of action of how LH stimulates Leydig cells to secrete testosterone occurs via activation of the cAMP second messenger system, thus increasing intracellular cAMP and activating protein kinase A, which results in phosphorylation of specific proteins, producing an increase in steroid production. Administration of CAJ for 42 days showed a significant decrease in LH levels in experimental Group II. The LH significant decrease is a reverse of what happened in FSH level, in which both are gonadotropins released from the anterior pituitary when stimulated by gonadotropin-releasing hormone (GnRH) from the hypothalamus. The decrease in LH in the CAJ-treated group could be a result of negative feedback on the anterior pituitary caused by an increase in testosterone in Group II [12]. An alternative explanation for the rise in FSH without an increase in LH could be related to the slight inhibition of spermatogenesis [16]. This is an indication that CAJ inhibits spermatogenesis, leading to a low level of LH, which in turn increases the level of testosterone, which activates cAMP second messengers and consequently causes protein phosphorylation for the production of steroids.

Testosterone was found to significantly increase in Group II Wistar rats that were given *cCAJ*. However, the LH significantly decreased, and since LH stimulates Leydig cells to release testosterone, it could, therefore, be said that the significant increase in testosterone had nothing to do with LH stimulation. The increase in testosterone levels can be attributed to the effect of the alkaloid component of CAJ, as previously reported by [17], which indicates that alkaloids are known as the starting material in the manufacture of steroid hormones. This indicates that the increase in testosterone levels in Group II might be triggered by alkaloids and thus could be because of their effect on the synthetic pathway of testosterone. Moreover, it could also be due to the probable effect of LH on the Leydig cells of the testes that causes an increase in the rate of conversion of cholesterol to pregnenolone and subsequently testosterone. The decrease in testosterone levels in Group III and Group IV could be due to decreases in alkaloid concentrations caused by the dilution ratio of the CAJ. Similarly, the same findings were reported by [14], who showed that *Citrus aurantifolia* caused a significant dose-dependent decrease in LH levels and a significant increase in testosterone levels using medium and high doses.

Prolactin is mostly found in nursing mothers in large quantities because of its role in lactation. Its action or contributions to spermatogenesis or testosterone secretion are not yet fully understood. Prolactin showed significant decreases in Group II compared with the normal control Group I. This effect may be due to the presence of flavonoids in the *cCAJ* as previously reported by [18] where flavonoid from *Vitex rotundifolia* was shown to inhibit prolactin production. However, the prolactin level significantly increased from III to IV when compared with Group II. The decreases in LH and prolactin and increases in FSH and testosterone in Group II and Group IV were found to be dependent on the concentration or dilution ratio of CAJ to water, as occurs with an increase in water. This is contrary to the finding of [14], who reported no effects of oral administration of CAJ at different concentrations on the serum prolactin levels of male Wistar rats.

The total sperm count, motility, viability and morphology are crucial indices that determine the potential of sperm to fertilize an ovum. Poor semen characteristics have been linked to damaged spermatocyte DNA [19]. It was reported that the integrity of spermatocyte DNA is protected by zinc and citric acid secreted from the prostate gland [20]. The mechanism involves neutralization of reactive oxygen species. In Group II, which received *cCAJ*, the mean values of total concentration, viable cells, AC, and normal cells were reduced significantly, and the mean values of sluggish cells, non-viable cells, non-motile cells, tail defect and head defect were also observed to have significantly increased relative to the normal control in Group I. However, the mean values significantly increased in Groups III and IV but were still lower than the normal control group. It is therefore most likely that as reported previously by [21], certain phytochemical constituents such as ganeisten, polyphenols and isflavon of CAJ might have either compromised the normal secretion of zinc and citric acid by the prostate or interrupted an enzymatic pathway involved in reactive oxygen species deactivation. Hence, semen parameters decrease in quantity and quality. However, the finding suggests changes in parameters as the concentration of the CAJ changes in ratios of 1:1 and 1:2 to water when compared with the *cCAJ*, which could be due to the increase in bioavailability of secondary metabolites such as alkaloids, saponins, flavonoids, tannins, and limonoids in the CAJ. Flavonoids in lime juice have been reported to exert mild antioxidative effects on lipid peroxidation [22]. Limonoids are highly oxygenated modified triterpenes derived from a precursor with a 4,4,8-trimethyl-17 furanyl steroid skeleton [23]. An increasing body of evidence seems to suggest that limonoids and flavonoids have independent biological activities. Alkaloids are also major components of stimulant agents such as

cocaine, caffeine and nicotine. Studies have shown that exposure of human sperm to nicotine *in vitro* decreases the percentage of motile sperm [22]. The alkaloid constituent has been implicated as responsible for this effect. The results obtained from this study strongly support that *Citrus aurantifolia* is a rich source of alkaloids, as demonstrated by other researchers [6, 14, 23, 24], and all suggested that the acidity of CAJ is suspected to be the reason for the destruction of sperm cells.

The dose-dependent reduction in sperm motility, percentage sperm concentration and percentage normal morphology indicate the anti-fertility potency of CAJ, which is directly or indirectly related to its physico-chemical characteristics and phytoconstituents. In a similar finding on oral administration of aqueous extract of *Aegle marmelos* to male rats, there was a significant decrease in the weights of reproductive organs. Sperm motility as well as sperm density in the cauda epididymis were also reduced significantly [24].

CONCLUSION

The effects of fresh juice of *Citrus aurantifolia* in male albino Wistar rats showed significant changes in all fertility indices in both groups that received cCAJ or dCAJ. The trend of these changes was also shown to be dependent on the degree of concentration of CAJ. The findings of this study revealed the negative effects of CAJ on male fertility factors that adversely affect spermatogenesis in male adult Wistar rats.

Disclaimer

The article has not been previously presented or published, and is not part of a thesis project

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgment

Authors acknowledge the contributions of Professor Zakari Muhammed of the Department of Obstetrics and Gynecology and Professor Sani Malami of Pharmacology Department, as well as staffs of Animal Laboratory, Bayero University Kano

REFERENCES

1. Isidori AM, Pozza C, Gianfrilli D, Isidori A. Medical treatment to improve sperm quality. *Reprod Biomed Online*. 2006 Jun;12(6):704-14. doi: 10.1016/s1472-6483(10)61082-6.
2. Cooper TG, Noonan E, von Eckardstein S, Auger J, Baker HW, Behre HM, Haugen TB, Kruger T, Wang C, Mbizvo MT, Vogelsong KM. World Health Organization reference values for human semen characteristics. *Hum Reprod Update*. 2010 May-Jun;16(3):231-45. doi: 10.1093/humupd/dmp048.
3. Oni PI. Ethnobotanical survey of a fallow plot for medicinal plants diversity in Idena village Ijebu-Ode, South-western Nigeria. *J Med Plants Res*. 2010 Apr 4;4(7):509-16.
4. Marieb EN, Hoehn K. Human anatomy & physiology. Pearson education; 2007.
5. Bowen R. Gonadotropins: luteinizing and follicle stimulating hormones. Colorado State University. 2004. Available at: <http://arbl.cvmbs.colostate.edu/hbooks/pathophys/endocrine/hypopit/lhfsh.html>
6. Hair WM, Gubbay O, Jabbour HN, Lincoln GA. Prolactin receptor expression in human testis and accessory tissues: localization and function. *Mol Hum Reprod*. 2002 Jul;8(7):606-11. doi: 10.1093/molehr/8.7.606.
7. Bole-Feysot C, Goffin V, Edery M, Binart N, Kelly PA. Prolactin (PRL) and its receptor: actions, signal transduction pathways and phenotypes observed in PRL receptor knockout mice. *Endocr Rev*. 1998 Jun;19(3):225-68. doi: 10.1210/edrv.19.3.0334.
8. Hameed S, Jayasena CN, Dhillon WS. Kisspeptin and fertility. *J Endocrinol*. 2011 Feb;208(2):97-105. doi: 10.1677/JOE-10-0265. Epub 2010 Nov 17.
9. Boulpaep EL, Boron WF. Male sex act. In: Boulpaep EL, Boron WF, eds. *Medical Physiology: A Cellular and Molecular Approach*. 1st ed. St. Louis, Mo: Elsevier Saunders: 2005; 1125.
10. Fox SI. Male reproductive system. In: Fox SI, eds. *Human Physiology*. 8th ed. Boston, MA: McGraw-Hill: 2004; 638-651.
11. Guyton A, Hall J. Body and control of the internal environment. In: Guyton A, Hall J, eds. *Textbook of Medical Physiology*. 12th ed. Philadelphia: Elsevier Inc.: 2006; 1003-1008.
12. de Kretser DM. Editorial: Is spermatogenic damage associated with Leydig cell dysfunction? *J Clin Endocrinol Metab*. 2004 Jul;89(7):3158-60. doi: 10.1210/jc.2004-0741.
13. Okon UA, Etim BN. Citrus aurantifolia impairs fertility facilitators and indices in male albino wistar rats. *Int J of Reprod Contraception*. 2014; 3:640-5.

14. Bakare AA, Basse RB, Okoko II, Sanyaolu AO, Ashamu AE, Ademola AO. Effect of lime juice (*Citrus aurantifolia*) on histomorphological alterations of the ovaries and uterus of cyclic Sprague-Dawley rats. *Europ J Sci Res.* 2012;67(4):607-16.
15. Fowler PA, Sorsa-Leslie T, Harris W, Mason. Ovarian gonadotrophin surge attenuating factor (GnSAF): where are we after 20 years of research? *Reprod.* 2010;126(6):689-99.
16. Ikpeme EV, Udoh PB, Ekaluo UB, Udensi O, Asuquo BO. Spermicidal and Hormonal response of wistar rats treated with ethanol seed, leaf and pulp extracts of *Carica papaya* (Linn). *Int J Recent Sci Res.* 2010;7:155-9.
17. YE Q, Zhang QY, Zheng CJ, Wang Y, Qin LP. Casticin, a flavonoid isolated from *Vitex rotundifolia*, inhibits prolactin release in vivo and in vitro. *Acta Pharmacol Sin.* 2010 Dec;31(12):1564-8. doi: 10.1038/aps.2010.178.
18. John Bray. Reproductive system. In: Bray J, Cragg P, Macknight A, Mills R, eds. *Lecture Notes on Human Physiology.* 4th ed. Hoboken, NJ: Wiley; 1999; 264-269.
19. Agarwal A, Said TM. Role of sperm chromatin abnormalities and DNA damage in male infertility. *Hum Reprod Update.* 2003 Jul-Aug;9(4):331-45. doi: 10.1093/humupd/dmg027.
20. Gonzales C, Leiva-Revilla J, Rubio J, Gasco M, Gonzales GF. Effect of red maca (*Lepidium meyenii*) on prostate zinc levels in rats with testosterone-induced prostatic hyperplasia. *Andrologia.* 2012 May;44 Suppl 1:362-9. doi: 10.1111/j.1439-0272.2011.01190.x.
21. Mankad M, Sathawara NG, Doshi H, Saiyed HN, Kumar S. Seminal plasma zinc concentration and alpha-glucosidase activity with respect to semen quality. *Biol Trace Elem Res.* 2006 May;110(2):97-106. doi: 10.1385/BTER:110:2:97.
22. Zini A, Libman J. Sperm DNA damage: importance in the era of assisted reproduction. *Curr Opin Urol.* 2006 Nov;16(6):428-34. doi: 10.1097/01.mou.0000250283.75484.dd.
23. Shamsi MB, Kumar R, Dada R. Evaluation of nuclear DNA damage in human spermatozoa in men opting for assisted reproduction. *Indian J Med Res.* 2008 Feb;127(2):115-23.
24. Chauhan OP, Archana BS, Singh A, Raju PS, Bawa AS. A refreshing beverage from mature coconut water blended with lemon juice. *J Food Sci Technol.* 2014 Nov;51(11):3355-61. doi: 10.1007/s13197-012-0825-6.