

Original article

Determination of Carbohydrate, Total Antioxidant, and Mineral Concentrations of *Linaria triphlla* (L) (Om lawlad), *Malva parviflora* Linn L. (Khabiza), and *Myrtus communis* L. (Birsim) Plants

Khadijah Al-Awjali¹, Mona Khanfar², Farag El-Mokasabi³, Zuhir Akrim⁴, Hamad Hasan⁵

¹Botany Department, Faculty of Science, Derna University, Libya

²Chemistry Department, Faculty of Science (Al-Quba), Derna University, Libya

³Botany Department, Faculty of Science, Benghazi University, Libya

⁴Pharmacology and Toxicology Department, Faculty of Pharmacy, Omar Al-Muhtar University, Libya

⁵Chemistry Department, Faculty of Science, Omar Al-Mukhtar University, Libya

Corresponding email. hamad.dr@omu.edu.ly

Abstract

This study investigated the biochemical composition of three plant species growing in northeastern regions of Libya: *Linaria triphlla* (Om lawlad), *Malva parviflora* (Khabiza), and *Myrtus communis* (Birsim). Leaves and stems were analyzed to determine antioxidant capacity, total phenol content, carbohydrate levels, and mineral concentrations (sodium, potassium, calcium). Spectrophotometry was used to estimate antioxidants, phenols, and carbohydrates, while flame photometry measured mineral content. Results revealed notable variations between leaves and stems. Total phenol content fluctuated between 194.61–295.45 ppm in leaves and 283.449–313.770 ppm in stems, indicating higher phenolic concentrations in stems. Antioxidant values showed minimal variation, ranging from 9.703–9.96 ppm in leaves and 10.198–10.528 ppm in stems. Carbohydrate levels were generally low, with values between 0.016–0.134 ppm in leaves and 0.155–0.419 ppm in stems. Mineral analysis demonstrated greater differences. Sodium concentrations ranged from 0.625–1.208 ppm in leaves and 1.88–16.88 ppm in stems, showing a marked increase in stems. Potassium levels were consistently higher in stems compared to leaves, while calcium concentrations were relatively small in leaves (0.12–0.68 ppm) but higher in stems (0.375–1.541 ppm). Overall, the study highlights those stems of the selected plants generally contain higher levels of phenols, minerals, and carbohydrates compared to leaves, while antioxidant capacity remains relatively stable across plant parts. These findings provide insight into the nutritional and biochemical properties of native Libyan plants, emphasizing their potential value in food science, pharmacology, and traditional medicine.

Keywords. Carbohydrate, Antioxidant, Phenols, Minerals, Plants, Libya.

Introduction

Around the world, the use of medicinal plants has taken center stage in health systems. This entails using medicinal plants not only to treat illnesses but also as a possible resource for preserving health and well-being. Due to its greater cultural acceptability, greater compatibility and flexibility with the human body, and less adverse effects, herbal medicine is used for primary healthcare in many nations, accounting for two-thirds of the global population. Based on documents, Plant extracts are found in the majority of medications. Some have bioactive compounds or active chemicals derived from plants. Despite recent advancements in science and technology, some of the claims and beliefs of indigenous people are irreplaceable. Recent research has led to the discovery of plant-derived drugs through the study of curative, therapeutic, traditional cures, and particularly the folk knowledge of indigenous people [1]. Aspirin, atropine, artimesinin, colchicine, digoxin, ephedrine, morphine, physostigmine, pilocarpine, quinine, quinidine, reserpine, taxol, tubocurarine, vincristine, and vinblastine are a few medications that are thought to be derived from plants. Examining the traits and functions of phytochemicals in some of the medical plants that are frequently utilized in Libya will be important since the significance of medicinal plants cannot be understated. Due to their potential to benefit humanity and society in many ways, particularly in the fields of health and pharmaceuticals, medicinal plants are receiving more attention than ever.

Because of its ethnochemical properties, plant parts including leaves, roots, and bark are employed for medicinal purposes and also act as precursors for the manufacture of beneficial pharmaceuticals. nature's medical significance [2]. The bioactive phytochemical components of these plants that have physiological effects on humans may be the source of their therapeutic potential. Due to their versatility, substances originating from plants have recently attracted a lot of attention. Phytochemicals are compounds found in plants that increase their global utility [3]. As a result, numerous investigations were conducted to predict a wide range of chemicals [4–38]. In addition, the amounts of metals and minerals were examined using various techniques in various samples [38–75]. The purpose of this study is to quantify some of the chemical components (antioxidants, total phenols, and carbohydrates) in a few chosen plants. utilizing the phytochemicals found in stems and leaves. To quantify the amounts of the minerals Na, K, and Ca in the leaves and stems of plants growing in the Al-Gabal Al-Ahder districts of Libya, namely *Linaria triphlla* (L) (Om lawlad), *Malva parviflora* Linn L. (Khabiza), and *Myrtus communis* L. (Birsim). in Libya's Al-Gabal Al-Ahder districts.

Methods

Sampling

The leaves and stems of *Linaria triphlla* (L) (Om lawlad), *Malva parviflora* Linn L. (Khabiza), and *Myrtus communis* L. (Birsim) plants were chosen for this study. Samples were gathered from various locations, such as the Wadi Derna valley and Karsah in the West, Al-Dhahr Al-Ahmar in the South, and the Mediterranean coast in the North. The study area is situated on the second terrace of El-Jabal El-Akhdar Mountain, which is situated in the Derna region of northeastern Libya. The city is divided into two sections by the Wadi, which runs between longitudes 33°00' and 32°30'N and 22°30' and 22°45'E. The Wadi is situated between 40 and 300 meters above sea level. With a mean temperature of roughly 20 °C, the study area's climate is similar to that of El-Jabal El-Akhdar. Rainfall often falls between 200 and 300 mm (Figure 1). Al-Dhahr Al-Ahmar in the south, the Mediterranean coast in the north, and Karsah in the west. The study area is situated on the second terrace of El-Jabal El-Akhdar Mountain, which is part of Wadi Derna in the Derna region of northeastern Libya. The Wadi separates the city into two halves between longitudes 33°00' and 32°30'N and 22°30' and 22°45'E. The Wadi is situated between 40 and 300 meters above sea level. With a mean temperature of roughly 20 °C, the study area's climate is similar to that of El-Jabal El-Akhdar. Rainfall often falls between 200 and 300 mm (Figure 1).

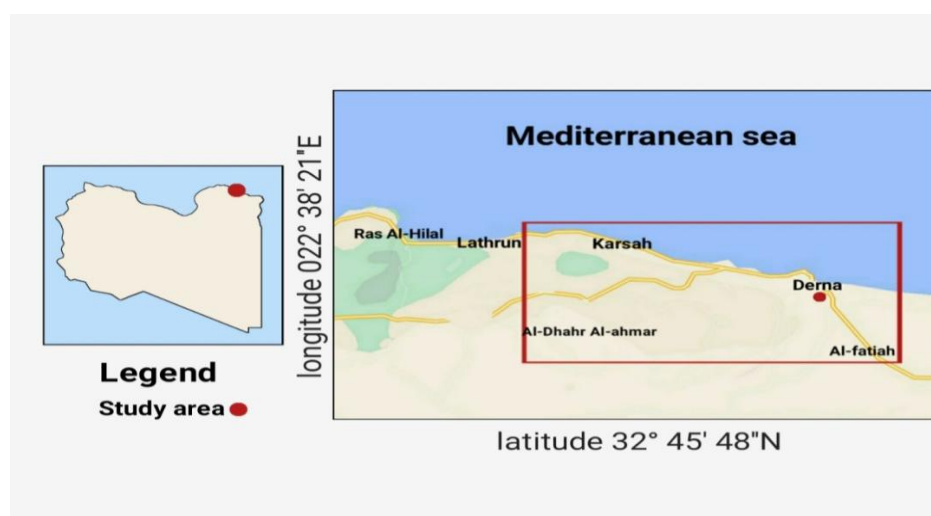


Figure 1. The studied area

Sample extraction

10 grams of each dried sample were taken and transferred to the beaker containing 100 ml of distilled water, and the mixture was mixed. Then the extraction was carried out by the evaporator system at 75 °C. After two hours, the mixture was filtered, and the filtrate was used to determine the phytochemical screening [6-10].

Determination of Phenol Compounds by Folin Ciocalteu Method

This experiment was carried out to determine Phenolic compounds, where the amount of total phenolic in the Extracts was determined by (the Folin-Ciocalteu) reagent according to the method of Slinkard and Singleton (10) using gallic acid as a standard. Samples (two replicates of the sample) were introduced into test cuvetts, then 1.0 ml of Folin-Ciocalteu reagent and 0.8 ml of Na₂CO₃ (7.5%) were added. The absorbance of all samples was measured at 765 nm using the Shimadzu UV – Vis spectrophotometer after incubating at 30 °C for 1.5 h. Results were expressed as ppm of fresh weight.

Determination of antioxidant capacity by the Prussian blue method

One gram of the powdered sample was defatted with petroleum ether. The defatted powder was then extracted sequentially by stirring with 10 ml of methanol twice, then extracted again with 10 ml 1% hydrochloric acid: methanol (v/v). The three combined extracts were evaporated under vacuum, and the residue was dissolved in 10 ml of methanol. Half ml of the solution was diluted with 3 mL of distilled water, 3 ml (0.008 M) of K₃Fe (CN)₆ was added, 3 ml 0.1M HCl, and 1 ml 1% FeCl₃. The blue color is allowed to develop for 5 minutes, and the absorbance is measured at 720 nm at the central lab of the Faculty of Science, Omar Al-Mukhtar University.

Determination of Carbohydrates

To estimate total carbohydrates, a known weight of 0.2 g of the dried sample was ground, then 5 ml of sulphuric acid was added. After completing the samples, the samples were cooled at room temperature, then a small quantity of Barium carbonate (Ba₂CO₃) was added, and the mixture was heated again. After cooling,

the samples were filtered. One ml of solution was taken, then one ml of 5% phenol was added. The total carbohydrate was determined by the method carried out in a previous study. Where the absorbance was measured at a wavelength of 490 nm [8-10].

Determination of Minerals

The sodium and potassium, calcium contents were measured by a Flame Photometer (JENWAY Flame Photometer) according to the method described by some studies [60-70]. In the central lab of the Faculty of Science, Omar Al-Mukhtar University.

Results

Total Phenols, Antioxidant, and Carbohydrate Contents

In this study, the concentrations of total phenols showed an increase in their values in stems of the studied plants compared with the leaves. The higher content (313.70 ppm), on the other hand, the lower content (194.61 ppm) was observed in the stems of *Myrtus communis* L. Sp. Pl plant. Generally, the contents of total phenols fluctuated as follows (from 194.61 to 295.45 ppm) and from 283.449 – 313.770 ppm) of leaves and stems, respectively, (Table 3). The antioxidant values ranged from 9.703 – 9.96 ppm in leaves and from 10.198 -10.528 ppm in stems. No wide variations were observed in the antioxidant contents of the studied plants. The carbohydrate contents did not show high values or high variations between leaves and stems of the studied plants. Where their values ranged between 0.016 and 0.134 ppm in leaves, and from 0.155 to 0.419 ppm for stems (Table 1).

Table 1. The contents (ppm) of Phenols Antioxidant and Carbohydrate in the studied samples

Scientific Name Compounds	Total Phenols		Antioxidant		Carbohydrate	
	Leaves	Stems	Leaves	Stems	Leaves	Stems
<i>Linaria Triphlla</i> (L) Mill Gard Dict	221.12	295.818	9.870	10.198	0.118	0.419
<i>Malva Parviflora</i> Linn. Demonstr. Pl	194.61	283.449	9.703	10.204	0.134	0.156
<i>Myrtus Communis</i> L. Sp. Pl.	295.45	313.770	9.96	10.528	0.016	0.155

Minerals

The results of this study recorded the presence of sodium, potassium and calcium in leaves and stems of the studied plants, the contents of sodium were ranged as following: (0.625 -1.208 ppm) and (1.88 -16.88 ppm) in leaves and stems, respectively, the results indicated that there as increase the sodium contents (16.8 ppm) in *Malva parviflora* Linn. Demonstr.Pl comparing with the other studied plants, especially in stems, the concentrations of potassium showed higher levels compared with sodium and calcium, also, there are high concentrations in *Malva parviflora* Linn. Demonstr.Pl plant compared with the other studied plants. The results showed an increase in the potassium concentrations in stems compared with leaf samples. Generally, the concentrations of potassium fluctuated in the ranges of 3.96 – 28.36 ppm in leaves and from 54.76 – 84.36 ppm in stems. The results also showed small amounts of calcium in leaves compared with stems, there is a relative increase of Calcium (1.541 ppm) in *Malva parviflora* Linn. Demonstr.Pl plant comparing with the other studied plants, (Table 2).

Table 2. The contents (ppm) of minerals (Na, K, and Ca) in the studied samples

Scientific Name Compounds	Sodium		Potassium		Calcium	
	Leaves	Stems	Leaves	Stems	Leaves	Stems
<i>Linaria Triphlla</i> (L) Mill Gard Dict	1.208	7.28	19.76	65.16	0.28	0.375
<i>Malva Parviflora</i> Linn. Demonstr. Pl	1.208	16.88	28.36	84.36	0.68	1.541
<i>Myrtus Communis</i> L. Sp. Pl.	0.625	1.88	3.96	45.76	0.12	0.708

Discussion

According to the study's findings, the plant being studied has tiny levels of carbohydrates, antioxidants, and total phenols. The existence of various phenol or flavonoid molecules, among others, is what primarily determines the antioxidant activity. The majority of plants that are categorized as medicinal typically contain several natural product chemicals that give them their effectiveness as antioxidants. Additionally, the antioxidant is linked to the presence of phenols in plants. An accurate equipment, such as GC-Mass or HPLC, which are used to identify the kinds and amounts of natural products or organic compounds, is required for the assessment of the types of phenols or flavonoids. The presence of carbohydrates indicates that the majority of the plant's secondary metabolites originate from various compounds, such as phenolic acids, tannins, terpenes, and carbohydrates; the slight variations in their contents are primarily caused by the physiological characteristics of each plant tissue [30–35].

The presence of sodium, potassium, and calcium minerals in leaves and stems was also noted in this study. The slight variations in these minerals' contents in the leaves and stems were also linked to the unique structure of each tissue. Additionally, the environmental conditions surrounding the plant sampling had a

major impact on the mineral contents in samples, including the types of soils, water, and the geochemical makeup of the plants growing [73–85]. The type of chemical analysis also has an impact on the amounts of minerals or metals in samples. A variety of instruments, such as atomic absorption, spectrophotometer, flame photometer, ionic coupling plasma, X-ray fluorescence, and others, can be used to estimate the metals in various samples, such as soil, water, plants, and others [86–95]. Some studies have also used the metals in other applications, such as antibacterial agents [96–103].

Conclusion

According to the results obtained in this study, the selected plants contain different natural compounds such as sterols, flavonoids, phenols, Alkaloids, Tannins, Anthraquinones, and saponins. Also, there are variations in their contents in leaves and stems. Small amounts of calcium were recorded compared with potassium and sodium. The results also showed that the studied plants contained different amounts of antioxidants and total phenols in both leaves and stems.

Acknowledgment

Special thanks to the staff members of the central laboratory of the chemistry department, Faculty of Science, Omar Al-Muhtar University, for their support during the establishment of this study.

Conflict

The authors state that no conflict in the results recorded in this study with other studies.

References

1. Vandebroek I, Calewaert J, De Jonckheere S, Sanca S, Semo L, Van Damme P, et al. Use of medicinal plants and pharmaceuticals by indigenous communities in the Bolivian Andes and Amazon. *Bull World Health Organ.* 2004;82(4):243-50.
2. Reyes-García V, Vadez V, Byron E, Apaza L, Leonard W, Perez E, et al. Market economy and the loss of ethnobotanical knowledge: estimates from Tsimane' Amerindians, Bolivia. *Curr Anthropol.* 2005;46(4):651-6.
3. Fagbohun ED, Lawal OU, Ore ME. The proximate, mineral and phytochemical analysis of the leaves of *Ocimum gratissimum* L., *Melanthera scandens* A. and *Leea guineensis* L. and their medicinal value. *Int J Appl Sci Technol.* 2012;2(1).
4. Al-Awjal K, Abdulsalam S, El-Mokasabi F, Akrim Z, Hasan H. Estimate the antioxidant capacity, total phenol contents, mineral concentrations, total carbohydrate of *Capparis spinosa* L. (Kabbar), *Ceratonia siliqua* L. (Kharuwab) and *Juniperus phoenicea* L. (Arar) plants. *Attahadi Med J.* 2025;2(4):376-84.
5. Abdull-Jalliel H, Sulayman A, Alhoreir M, Hasan H. Antimicrobial effect of some metal concentration on growth of *Staphylococcus* and *Klebsiella* bacteria species. *AlQalam J Med Appl Sci.* 2025;8(3):1646-56.
6. Abdull-Jalliel H, Arous NB, Alhoreir M, Hasan H. Using extracts of Dodder plant and concentrations of some metals as inhibitors for growth of *Pseudomonas* bacteria isolated from some hospital rooms in Derna and Al bayda. *AlQalam J Med Appl Sci.* 2025;8(3):1600-11.
7. Eltawaty SA, Abdalkader GA, Hasan HM, Houssein MA. Antibacterial activity and GC-MS analysis of chloroform extract of bark of Libyan *Salvia fruticosa* Mill. *Int J Multidiscip Sci Adv Technol.* 2021;1(1):715-21.
8. Aljamal MA, Hasan HM, Al Sonosy HA. Antibacterial activity investigation and anti-biotic sensitive's for different solvents (Ethanol, propanol, DMSO and diethyl ether) extracts of seeds, leaves and stems of (*Laurus azorica* and *Avena sterilis*) plants. *Int J Curr Microbiol Appl Sci.* 2024;13(11):175-90.
9. Hamade MH, Abdelraziq SA, Gebreel AA. Extraction and determination of Beta carotene content in carrots and tomato samples collected from some markets at ElBeida City, Libya. *EPH Int J Appl Sci.* 2019;1(1):105-10.
10. Hasan HM, Ibrahim H, Gonaïd MA, Mojahidul I. Comparative phytochemical and antimicrobial investigation of some plants growing in Al Jabal Al-Akhdar. *J Nat Prod Plant Resour.* 2011;1(1):15-23.
11. Hasan H, Jadallah S, Zuhir A, Ali F, Saber M. Anti-Cancer, Anti-Inflammatory, Antibacterial, Antifungal, Anti-Oxidant and phytochemical investigation of flowers and stems of *Anacyclus clavatus* plant extracts. *AlQalam J Med Appl Sci.* 2025;8(3):415-27.
12. Hasan H, Zuhir A, Shuib F, Abdraba D. Phytochemical investigation and exploring *Citrullus Colocynthis* extracts as antibacterial agents against some gram and negative bacteria species. *AlQalam J Med Appl Sci.* 2025;8(3):392-400.
13. Zeyaullah MD, Naseem A, Badrul I, Hamad MI, Azza SA, Faheem AB, et al. Catechol biodegradation by *Pseudomonas* strain: a critical analysis. *Int J Chem Sci.* 2009;7(3):2211-21.
14. El-Mehdawy MF, Eman KS, Hamad MI, Hasan H. Amino acids contents of leaves and stems for two types of herbal plants (Marjoram and Hybrid tea rose) at AL-Gabal AL-Akhder region. *Der Pharma Chem.* 2014;6(6):442-7.
15. El-Mehdawy MF, Eman KS, Hamad MIH. Amino acid contents of leaves and stems for three types of herbal plants at AL-Gabal AL-Akhder region. *World J Chem.* 2014;9(1):15-9.
16. Hamad MH, Noura AAM, Salem AM. Phytochemical screening, total phenolic, anti-oxidant, metal and mineral contents in some parts of *Plantago albicans* grown in Libya. *World J Pharm Res.* 2024;13(3):1-17.
17. Anees AS, Hamad MIH, Hasan H, Mojahidul I. Antifungal potential of 1,2,4-triazole derivatives and therapeutic efficacy of *Tinea corporis* in albino rats. *Der Pharm Lett.* 2011;3(1):228-36.
18. Hasan H, Mohammed M, Haroon A. Determining contents of antioxidants, total phenols, carbohydrate, total protein, and some elements in *Eucalyptus gomphocephala* and *Ricinus communis* plant samples. *Libyan Med J.* 2015;1(1):222-31.

19. Hasan H, Akrim Z, Shuib F, Abdraba D. Efficiency of *Cynara Cornigera* fruits on antibacterial, antifungal and its phytochemical, anti-oxidant screening. *Libyan Med J.* 2025;3(1):120-8.
20. Hasan H, Sulayman A, Alehrir A. Estimation of amino acid composition, total carbohydrate, and total protein content in *Ballota pseudodictamnus* plant extracts from Al Jabal Al Akhdar Region, Libya. *Libyan Med J.* 2025;3(1):266-71.
21. Hasan H, Hamad A, Abdelsatar W. Evaluation of anti-oxidant capacity, total phenol, metal, and mineral contents of *Ziziphus lotus* plant grown at some regions of AlGabal AlKhder, Libya. *Libyan Med J.* 2025;3(1):137-43.
22. Ben Arous NA, Naser ME, Hamad MAH. Phytochemical screening, anti-bacterial and anti-fungi activities of leaves, stems and roots of *C. parviflorus* Lam and *C. salviifolius* L plants. *Int J Curr Microbiol App Sci.* 2014;13(11):262-80.
23. Anas FAE, Hamad MAH, Salim AM, Azza MH. Phytochemical screening, total phenolics, antioxidant activity and minerals composition of *Helichrysum stoechas* grown in Libya. *Afr J Biol Sci.* 2024;3(6):2349-60.
24. Naseer RE, Najat MAB, Salma AA, Hamad MAH. Evaluation of metal and mineral contents of leaves, stems and roots of *C. Parviflorus* Lam and *C. Salviifolius* L plants growing at Al Ghabal Al-Khder (Libya). *Int J Adv Multidiscip Res Stud.* 2024;4(5):191-4.
25. Hamad MAH, Salem AM. Total carbohydrate, total protein, minerals and amino acid contents in fruits, pulps and seeds of some cultivars of muskmelon and watermelon fruit samples collected from AlGabal Alkhder region. *Sch J Appl Med Sci.* 2024;12(1):1-7.
26. Gonaïd MI, Ibrahim H, Al-Arefy HM. Comparative chemical and biological studies of *Salvia fruticosa*, *Ocimum basillicum* and *Pelargonium graveolans* cultivated in Al-Jabal Al- Akhdar. *J Nat Prod Plant Resour.* 2012;6(2):705-10.
27. Rinya FMA, Hamad MAH, Ahlam KA, Hammida MEH. Phytochemical screening of some herbal plants (*Men, Origanum* and *Salvia*) growing at Al-Gabal Al-akhder Region-Libya. *Afr J Basic Appl Sci.* 2017;9(3):161-4.
28. Anas FAA, Hamad MAH, Salim AA, Azza MH. Phytochemical screening, total phenolics, antioxidant activity and minerals composition of *Helichrysum stoechas* grown in Libya. *Afr J Biol Sci.* 2024;3(6):2349-60.
29. Haroon A, Hasan H, Wafa AAS, Baset ESM. A comparative study of morphological, physiological and chemical properties of leaves and steam samples of (*E.gomphocephala*) (Tuart) plant growing at coastal (Derna city) and *J Res Environ Earth Sci.* 2024;9(12):10-8.
30. Hamad MAS, Ali AR. Separation and identification speciation of phenolic compounds in fruits and leaves of some medicinal plants (*Juniperus phoenicea* and *Quercus coccifera*) growing at AlGabal Al Akhdar region, Libya. *Indian J Pharm Educ Res.* 2016;51(3):299-303.
31. Enam FM, Wesam FAM, Hamad MAH. Detection contents of minerals of (Sodium, Potassium and Calcium) and some metals of (Iron, Nickel and Copper) in some vegetable and the samples collected from Al-Marj. *Int J Adv Multidiscip Res Stud.* 2023;5(3):304-9.
32. Hamad MIH, Mousa SR. Synthesis and (IR and TEM) characterization of leaves and stem nanoparticles of *Artemisia* plant: comparative study for evaluation of anti-bacterial efficiency. *Int J Adv Multidiscip Res Stud.* 2024;4(5):195-9.
33. Elsalhin H, Abobaker HA, Hasan H, El-Dayek GA. Antioxidant capacity and total phenolic compounds of some algae species (*Anabaena* and *Spirulina platensis*). *Sch Acad J Biosci.* 2016;4(10):782-6.
34. Alaila AK, El Salhin HE, Ali RF, Hasan HM. Phytochemical screening of some herbal plants (*Men, Origanum* and *Salvia*) growing at al-gabal al-akhder region- Libya. *Int J Pharm Life Sci.* 2017;8(4):5500-3.
35. Hasan H, Mariea FFE, Eman KS. Contents of some chemical compounds of leaves and stems of some herbal plants (*Thymy, Rosemary, Salvia, Marjoram* and *Hybrid Tea Rose*) at Al-Gabal Al-Akhder region. *EPH Int J Appl Sci.* 2014;6(3):1-8.
36. El-Mehdawe MF, Eman KS, Hamad MIH. Heavy metals and mineral elements contents of leaves and stems for some herbal plants at AL-Gabal AL-Akhder region. *Chem Sci Rev Lett.* 2014;3(12):980-6.
37. Hamad MIH, Aaza IY, Safaa SHN, Mabrouk MS. Biological study of transition metal complexes with adenine ligand. *Proc.* 2019;41(1):77.
38. Hasan JA, Hasan HMA. Potential human health risks assessment through determination of heavy metals contents in regularly consumed yogurta in Libya. *World J Pharm Pharm Sci.* 2024;13(12):100-12.
39. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Essam AM, Hamad MIH. Rice husk and activated carbon for waste water treatment of El-Mex Bay, Alexandria Coast, Egypt. *Arab J Chem.* 2016;9(S2):S1590-6.
40. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Hamad IH. Heavy metals accumulation in sediments of Alexandria coastal areas. *Bull Fac Sci.* 2012;47(1-2):12-28.
41. Mamdouh SM, Wagdi ME, Ahmed MA, Hamad MIH. Chemical studies on Alexandria coast sediment. *Egypt Sci Mag.* 2005;2(4):93-102.
42. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Hamad MIH. Distribution of different metals in coastal waters of Alexandria, Egypt. *Egypt Sci Mag.* 2010;7(1):1-19.
43. Mohamed AE, Afnan SA, Hamad MA, Mohammed AA, Mamdouh SM, Alaa RE, et al. Usage of natural wastes from animal and plant origins as adsorbents for removal of some toxic industrial dyes and heavy metals in aqueous media. *J Water Process Eng.* 2023;55:104192.
44. Mohamed HB, Mohammed AZ, Ahmed MD, Hamad MAH, Doaa AE. The heavy metal pollution and associated toxicity risk assessment in Ajdabiya and Zueitina, Libya. *Sci J Damiatta Fac Sci.* 2024;14(1):16-27.
45. Nabil B, Hamad H, Ahmed E. Determination of Cu, Co and Pb in selected frozen fish tissues collected from Benghazi markets in Libya. *Chem Methodol.* 2018;2:56-63.
46. Wesam FAM, Hamad MAH. Detection of heavy metals and radioactivity in some bones of frozen chicken samples collected from Libyan markets. *Int J Adv Multidiscip Res Stud.* 2023;3(3):761-4.
47. Wesam FAM, Hamad MAH. Study accumulation of minerals and heavy metals in *Ulva* algae, *Cladophora*, *Polysiphonia* and *Laurencia* algae samples at eastern north region of Libya coast. *GSC Biol Pharm Sci.* 2023;23(03):147-52.

48. Citrine E, Hamad H, Hajer Af. Contents of metal oxides in marine sediment and rock samples from eastern Libyan coast, utilizing X-ray method. *AlQalam J Med Appl Sci.* 2015;1(1):1316-21.
49. Hanan MA, Hamida E, Hamad MAH. Nitrogen, phosphorus and minerals (Sodium, Potassium and Calcium) contents of some algae's species (*Anabaena* and *Spirulina platensis*). *Int J Curr Microbiol App Sci.* 2016;5(11):836-41.
50. Mardhiyah F, Hamad H. Assessment of the contamination by heavy metals in Al-Fatayeh Region, Derna, Libya. *AlQalam J Med Appl Sci.* 2025;8(3):1081-91.
51. Abdelrazeg A, Khalifa A, Mohammed H, Miftah H, Hamad H. Using melon and watermelon peels for removal of some heavy metals from aqueous solutions. *AlQalam J Med Appl Sci.* 2025;8(3):787-96.
52. Abdul Razaq A, Hamad H. Estimate contents and types of water well salts by Palmer Roger model affecting corrosion of Al-Bayda city (Libya) network pipes. *AlQalam J Med Appl Sci.* 2025;8(3):744-53.
53. Abdulsayid FA, Hamad MAH, Huda AE. IR spectroscopic investigation, X-ray fluorescence scanning, and flame photometer analysis for sediments and rock samples of Al-Gabal Al-Akhder coast region (Libya). *IOSR J Appl Chem.* 2021;14(4):20-30.
54. ALambarki M, Hasan HMA. Assessment of the heavy metal contents in air samples collected from the area extended between Albayda and Alquba cities (Libya). *AlQalam J Med Appl Sci.* 2025;8(3):695-707.
55. Al-Nayyan N, Mohammed B, Hamad H. Estimate of concentrations of heavy metals in the and some plant samples collected from (near and far away) of main road between Al-Bayda city and Wadi Al-Kouf region. *AlQalam J Med Appl Sci.* 2025;8(3):816-26.
56. Hasan HMI. Studies on physicochemical parameters and water treatment for some localities along coast of Alexandria [Doctoral dissertation]. [Alexandria, Egypt]: Alexandria University; 2006.
57. Hamad MAH, Hager AA, Mohammed EY. Chemical studies of water samples collected from area extended between Ras Al-Halal and El Haniea, Libya. *Asian J Appl Chem Res.* 2022;12(3):33-46.
58. Hamad M, Mohammed AA, Hamad MAH. Adsorption and kinetic study for removal some heavy metals by use in activated carbon of sea grasses. *Int J Adv Multidiscip Res Stud.* 2024;4(6):677-85.
59. Hamad MAH, Hamad NI, Mohammed MYA, Hajir OAA, Al-Hendawi RA. Using bottom marine sediments as environmental indicator state of (Tolmaitha – Toukra) region at eastern north coast of Libya. *Sch J Eng Tech.* 2024;2(14):118-32.
60. Hamad MIH. Heavy metals distribution at coastal water of Derna city (Libya). *Egypt J Aquat Res.* 2008;34(4):35-52.
61. Hamad MIH, Mojahid ul Islam. Concentrations of some heavy metals of Al-Gabal Al-Akhder Coast Sediment. *Arch Appl Sci Res.* 2010;2(6):59-67.
62. Hamad MAH, Amira AKA. Estimate concentrations of some heavy metals in some shoes polish samples. *EPH Int J Appl Sci.* 2016;2(2):24-7.
63. Hamad MAH, Hussien SSM, Basit EEM. Accumulation of some heavy metals in green algae as bio indicators of environmental pollution at Al-Haniea region: Libya coastline. *Int J Adv Multidiscip Res Stud.* 2024;4(5):188-90.
64. Hamad MIH, Ahmed MA. Major cations levels studies in surface coastal waters of Derna city, Libya. *Egypt J Aquat Res.* 2009;35(1):13-20.
65. Hamad MIH, Masoud MS. Thermal analysis (TGA), diffraction thermal analysis (DTA), infrared and X-rays analysis for sediment samples of Toubrouk city (Libya) coast. *Int J Chem Sci.* 2014;12(1):11-22.
66. Hamad R, Ikraiam FA, Hasan H. Estimation of heavy metals in the bones of selected commercial fish from the eastern Libyan coast. *J Rad Nucl Appl.* 2024;9(1):47-51.
67. Hasan HAH. Estimate lead and cadmium contents of some archeological samples collected from ancient cities location (Cyrene and Abolonia) at Al-Gabal Al-Akhder Region, Libya. *Univ J Chem Appl.* 2021;12(21):902-7.
68. Hasan H, El-maleh C. Evaluation of some heavy metal levels in tissue of fish collected from coasts of susa region, libya. *Attahadi Med J.* 2025;1(1):118-22.
69. Balal A, Obid M, Khatab H, Hasan H. Determination of lead and cadmium marine water and crabs (*pachygrapsus marmoratus*) from tolmitha coast, Libya. *AlQalam J Med Appl Sci.* 2025;8(3):1670-7.
70. Arora D, Kaur J. Antimicrobial activity of spices. *Int J Antimicrob Agents.* 1999;12(3):257-62.
71. Bhat KS. Medicinal and plant information databases. In: *Medicinal Plants for Forests Conservation and Health Care.* FAO; 2011:158.
72. Anulika NP, Ignatius EO, Aymond ES, Osasere OL, Abiola AH. The chemistry of natural product; Plant secondary metabolites. *Int J Technol Emerg Eng Res.* 2016;4(8):1-9.
73. Hamad IH, Nuesry MS. Poly cyclic hydrocarbons levels in some fishes tissues collected from Derna City (Libya) Coast. In: *International conference on chemical, agricultural and medical sciences; 2014 Dec 4-5; Antalya, Turkey; 2014.* p. 52-6.
74. Hamad MAH, Mounera AAE, Baseet ESM, Eman E, Al-Badri M. Identification and detection aromatic and aliphatic hydrocarbons in *Epinephelus Marginatus* fish samples collected from Benghazi coast. *Int J Adv Multidiscip Res Stud.* 2023;6(3):107-13.
75. Mohammed A, Hamad MAH, Mounera AAE, Eman IHE. Extraction and identification of aliphatic hydrocarbons in marine sediment samples at Benghazi city and Dyriana town coasts (Libya). *J Res Humanit Soc Sci.* 2023;11(10):168-74.
76. Hasan MAH, Muftah HS, Abdelghani KA, Saad SI. Poly aromatic hydrocarbon concentrations in some shell samples at some Tobrouk city coast regions: could the oil industry be significantly affecting the environment. *Ukr J Ecol.* 2022;12(3):21-8.
77. Habel AMA, Mohamed NIH, Mohammed MA, Hamad MAH. Levels and sources of aliphatic and polycyclic aromatic hydrocarbons in blue runner fish from Benghazi coast, Libya. *Afr J Biol Sci.* 2024;6(3):1-10.
78. Hasan HMI, Mohamad ASA. A study of aliphatic hydrocarbons levels of some waters and sediments at Al-Gabal Al-Akhder coast regions. *Int J Chem Sci.* 2013;11(2):833-49.

79. Salem GM, Aljidaemi FF, Hwisa SA, Hamad MIH, Zaid AA, Amer IO. Occupational exposure to benzene and changes in hematological parameters in East Tripoli, Libya. *Nanotechnol Percept*. 2024;20(S5):358-64.
80. Habil Z, Ben arous N, Masoud N, Hasan H. Using GC-mass method for determination hydrocarbon compounds in some vegetable samples at Derna city, Libya. *Libyan Med J*. 2025;17(3):374-83.
81. Hasan H, Habil Z, Ben arous N. Estimate the types and contents of phenolic acid in *C.Paviflorus lam* and *C.salviifolius L* plants growing at Al –Gabal Al-hder regions. *AlQalam J Med Appl Sci*. 2025;8(3):1646-56.
82. Zeyadah MA, Bahnasaway MH, Deedah AM, El-Emam DA, Hamad MA Hasan. Evaluation of the contents of aliphatic and aromatic hydrocarbons in sediment from Zwwitina harbor coast (Libya), an indicator of petroleum pollution. *Egypt J Aquat Biol Fish*. 2023;27(6).
83. Hamad R, Ikraiam F, Hasan H. Determination of specific natural radionuclides in the bones of some local fish commonly consumed from the eastern Libyan coast. *J Rad Nucl Appl*. 2023;8(3):283-9.
84. Sroor AT, Walley El-Dine N, El-Bahi SM, Hasa HMA, Ali JM. Determination of radionuclides levels and absorbed dose for the, rock, plant and water in gondola- Libya. *IOSR J Appl Phys*. 2018;10(4):40-9.
85. Hasan H, Ammhmmid R, Khatab H, Ali J, Al kaseh A. Using gamma ray radiation to estimate the types and contents of radioactive nuclides in some ported sugar samples, Libya. *AlQalam J Med Appl Sci*. 2025;8(3):1795-803.
86. Hasan H, Abdelgader I, Emrayed H, Abdel-Gany K. Removal of the medical dye safranin from aqueous solutions by sea grasses activated carbon: a kinetic study. *AlQalam J Med Appl Sci*. 2025;8(3):428-34.
87. Hasan HMA, Alhamdy MA. Adsorption and kinetic study for removal some heavy metals by using activated carbon of sea grasses. *Int J Adv Multidiscip Res Stud*. 2024;4(6):677-85.
88. Almadani EA, Hamad MAH, Kwakab FS. Kinetic study of the adsorption of the removal of bromo cresol purple from aqueous solutions. *Int J Res Granthaalayah*. 2019;7(12):1-10.
89. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Essam AM, Hamad MIH. Rice husk and activated carbon for waste water treatment of El-Mex Bay, Alexandria Coast, Egypt. *Arab J Chem*. 2016;9(S2):S1590-6.
90. Alfutisi H, Hasan H. Removing of thymol blue from aqueous solutions by pomegranate peel. *EPH Int J Appl Sci*. 2019;1(1):111-9.
91. Ahmed ONH, Hamad MAH, Fatin ME. Chemical and biological study of some transition metal complexes with guanine as ligand. *Int J New Chem*. 2023;10(3):172-83.
92. Hamad MAH, Enas UE, Hanan AK, Hana FS, Somia MAE. Synthesis, characterization and antibacterial applications of compounds produced by reaction between Barbitol with Threonine, glycine, lycine, and alanine. *Afr J Biol Sci*. 2024;6(4):1-10.
93. Emrayed H, Hasan H, Liser R. Corrosion inhibition of carbon steel using (Arginine –levofloacin-metal) complexes in acidic media. *AlQalam J Med Appl Sci*. 2025;8(3):1633-40.
94. Hasan S, Abduljalil O, Mohamed F, Hasan H. Detection of residual pesticides (Imidacloprid ,Aldicarb,Metalaxyl,Cypermethrin ,Chlorpyrfos,DDA, and Endrin) in peach Samples collected from Jabal al Akhder farma,Libya. *AlQalam J Med Appl Sci*. 2025;8(4):2099-2106.
95. Mohamed FH, Salah MIH, Omuthum A, Hasan Hamad. Sensitive and rapid method to estimate residual pesticides in some local and imported apple cultivars collected from eastern north side of Libya. *Int J Adv Multidiscip Res Stud*. 2023;3(6):100.
96. Hamad MIH, Yahya Al, Hassan SS, Salama MM. Biological study of transition metal complexes with adenine ligand. *Proceedings*. 2019;41(1):77.
97. Habib IH, Idres AMA, Hasan HMI. Synthesis, Infrared (I.R), Thermal Gravimetric Analysis (TGA) Characterization and Antibacterial Activity of some Amino Acids Complexes. *Chem Sci Rev Lett*. 2014;3(12):1303-16.
98. Hasan HMA, Khalid HAA, Abdulsayid F. Infrared (IR) characterizations and physicochemical properties of Schiff base compound obtained by the reaction between 4-hydroxy-3-methoxy benzaldehyde and 2-amino-3-methylbutanoic acid. *J Res Pharm Sci*. 2021;7:8-12.
99. Salam MM, Moussa SF, Hasan HA. Preparation, Characterization and Antibacterial Activities of Metal complexes with Tyrosine Ligands. *Egypt J Chem*. 2023;66(10):1-7.
100. Zawia A, Najar AM, Abdusalam AAA, Aeyad T. Synthesis, Characterization, Biological Screening and Molecular Docking of New Schiff Base and Its Mononuclear Complexes with Pb²⁺, Cd²⁺, Zn²⁺ and Cu²⁺. *J Pharm Appl Chem*. 2023;8(1):1-9.
101. Hasan HMA, Elmagbari F, Othman A, Hammouda AN. Chemical and Biological Study of Some Transition Metal Complexes with Guanine as Ligand. *Int J New Chem*. 2023;10(3):172-83.
102. Hasan HMA, Al-Warad Y. Synthesis, physical properties, Infrared (IR) analysis and anti-fungi activity of some Valine and metal ion Complexes. *Int J Multidiscip Sci Adv Technol*. 2021;1:246-54.
103. Alfutisi HMM, Hasan HMI. Removing of Thymol Blue from aqueous solutions by Pomegranate peel. *EPH Int J Appl Sci*. 2019;1(1):111-9.