

Original article

Using Ionic Coupled Plasma (ICP) for estimating some Heavy Metals and elements in some Tea Samples and calculating their Health Risk Assessment

Hamad Hasan^{*1}, Marwa Khalifa², Monira Saleh¹

¹Chemistry Department, Faculty of Science, Omar Al-Mukhtar University, Libya

²Higher Institute of Medical Sciences and Technology, Al-Marj, Libya

Corresponding Email: hamad.dr@omu.edu.ly

Abstract

Tea is one of the most widely consumed beverages globally, including in Libya. This study investigates the concentrations of heavy metals in twelve tea samples available in Libyan markets using Inductively Coupled Plasma Mass Spectrometry (ICP-MS). The health risks associated with these metals were evaluated using the Target Hazard Quotient (THQ) and Hazard Index (HI). The results of this study recorded that some metals were detected in the studied Tea samples, including: Fe, Zn, Mn, Al, Cu, Cr, Ni, Ca. On the other hand, the metals of Pb, Co, Cd, Ag, As, and Sb were not detected in the studied samples. The contents of detected metals were fluctuated in the ranges of (0.24 – 1.08), (0.22 – 2.85), (1.68 – 6.69), (1.80 – 6.02), (0.060 – 0.13), (0.010 – 0.023), (ND – 0.022) and (16.97 – 27.87 µg/g), for Fe, Zn, Mn, Al, Cu, Cr, Ni and Ca, respectively. The higher concentrations were obtained for Mn, followed by Cr, then Fe. Results showed elevated levels of manganese (Mn) exceeding the Maximum Residue Limits (MRLs), while other metals such as zinc (Zn), iron (Fe), aluminum (Al), copper (Cu), chromium (Cr), nickel (Ni), and calcium (Ca) were within acceptable ranges. However, THQ and HI values for Mn and Cr in some samples exceeded safe thresholds, indicating potential health risks. These findings highlight the need for stricter quality control and monitoring of imported tea products in Libya.

Keywords: Heavy Metals, Tea, ICP-MS, Health Risk Assessment, Libya, THQ, HI

Introduction

Tea, a widely consumed beverage second only to water, plays a significant role in maintaining intestinal health by influencing the gut microbiota and bolstering immunity against intestinal disorders. Additionally, tea possesses antioxidant properties that protect cell membranes from oxidative damage. However, the cultivation of tea plants can be susceptible to various diseases and pests. To mitigate these challenges and ensure optimal quality, farmers often rely on pesticides [1]. Pesticides, although crucial for pest control and plant health, can have adverse effects on human health when used excessively. The cultivation, processing, and storage of tea leaves, especially black tea, can result in the accumulation of heavy metals. The World Health Organization (WHO) has established stringent guidelines for the permissible levels of heavy metals in tea. To guarantee the safety of consumers, rigorous monitoring of these substances in tea samples sourced from both tea gardens and commercial brands is imperative. This research endeavors to quantify the concentration of heavy metals in tea samples and evaluate the associated health risks. By conducting a comprehensive analysis, we aim to contribute to ensuring the safety and quality of tea products available to consumers.

The concentration of different metal components in tea can vary significantly, potentially impacting human health [2]. The accumulation of heavy metals in tea leaves can be attributed to both natural factors, such as soil contamination, and anthropogenic factors, such as the use of pesticides and fertilizers [3]. While some trace metals, like chromium, iron, cobalt, nickel, and zinc, are essential for normal physiological functions, other heavy metals, such as Pb, Ca, and arsenic, are not only non-essential but also toxic [4]. A critical biological characteristic of metals is their propensity for bioaccumulation. Consequently, bioaccumulation is a key factor in risk assessment strategies. For example, the calculation of the available percent of Aluminum (Al) and Zinc (Zn) in tea consumed by humans revealed that tea can contribute up to 37.2% of the daily dietary intake of Al, while only a small fraction, approximately 1.78%, is absorbed by the intestines [5].

Inductively coupled plasma mass spectrometry (ICP-MS) is a powerful technique for analyzing analytes that employs an inductively coupled plasma to ionize a sample. This process atomizes the sample, generating atomic and small polyatomic ions, which are subsequently detected. ICP-MS is renowned for its capability to detect trace metals and various non-metals in liquid samples with exceptional sensitivity. Furthermore, it can differentiate between isotopes of the same element, making it a versatile tool for isotopic labeling studies [6]. ICP-MS offers superior speed, precision, and sensitivity compared to atomic absorption spectroscopy. However, compared to other mass spectrometry techniques such as TIMS and GD-MS, ICP-MS introduces various interfering species, including argon from the plasma, air contaminants, and impurities from glassware and cones [7].

Tea is a staple beverage in Libyan households, consumed daily across all age groups. However, tea leaves can accumulate heavy metals from soil, water, and atmospheric deposition. Chronic exposure to these

metals poses significant health risks. The detection of chemical compounds was carried out on different samples in Libya such as hydrocarbon compounds [8-14], radioactive elements [15-19], or plants and their applications [20-55]. The metals have importance if they are used as complexes for different uses, but they also have other negative effects on the environment, different studies were carried out on different samples [56-94]. This study aims to quantify heavy metal concentrations in tea samples and assess the associated health risks.

Methods

Sampling

Twelve tea samples were collected from various Libyan markets. The samples are given in (Table 1).

Table 1. The studied Tea samples

Sample No	Sample Name	TeaType
1	Alagzalin	Black Tea
2	Ahmed Tea	
3	Zahra	
4	AlSafina	
5	Al wadg	
6	Nesma	
7	AlBaraka	Green Tea
8	AlgzaLin	
9	AlNajee	
10	AlBaraka	
11	Al Nabt	
12	Nessma	

Heavy metal determination

Samples were digested using concentrated nitric acid (HNO₃). 0.5 grams of each sample was added to a beaker containing 50 ml of distilled water, then 5 ml of concentrated nitric acid was added. The mixture was heated on a hot plate until near dryness, then cooled and filtered. The filtrate was brought to a volume of 100 ml in a measuring flask with distilled water (60-61). and analyzed using ICP-MS. Health risk assessment was conducted using THQ and HI formulas based on daily intake and reference doses.

Risk assessment was also done by calculating the target risk quotient (THQ), designed by the US Environmental Protection Agency which was designed by the US Environmental Protection Agency to determine safe levels of frequent long-term exposure to chemical pollutants. The THQ is a relationship between the measured concentration and the oral reference dose, weighted by the length and frequency of exposure, amount ingested, and body weight. A THQ reading of 1 or above indicates a health risk.

$$\text{THQ} = ([\text{Efr} * \text{ED tot} * \text{IFR} * \text{C}]) / ([\text{RfDo} * \text{BWa} * \text{ATn}]) * 10^{-3}$$

Where:

Efr: the exposure frequency (365 days/year); EDtot: is the exposure duration (80 years for the Italian population; it was the same for the Pakistani population as determined from survey results) according to ISTAT 2013. IFR: is the food ingestion rate (g day⁻¹); C: is the concentration (µg g⁻¹); RfDo: is the oral reference dose (µg g⁻¹ day⁻¹). BWa: is the adult body weight (60 kg). ATn: is the average time for non-carcinogens (it is equal to Efr × EDtot). The health risk index imposes a level of risk due to the presence of pesticides and metals in tea samples.

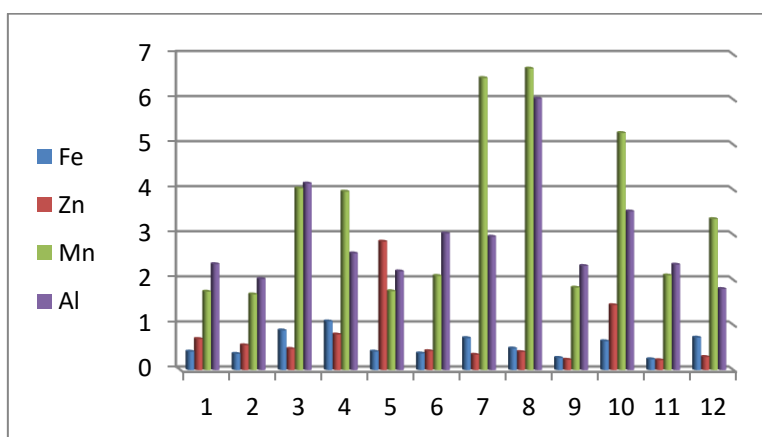
Results

Heavy metal contents

Some of heavy metals were detected in this study in tea samples by I.C.P, The metals including: Fe , Zn , Mn, Al, Cu, Cr, Ni, Ca, On the other side the metals of pb, Co, Cd, Ag, As and Sb were not detected in the studied samples, the contents were given in the (Tables 2-3) and (Figures 1-2). The contents of detected metals were fluctuated in the ranges of (0.24 – 1.08), (0.22 – 2.85), (1.68 – 6.69), (1.80 – 6.02), (0.060 – 0.13), (0.010 – 0.023), (ND – 0.022) and (16.97 – 27.87 µg/g), for Fe, Zn, Mn, Al, Cu, Cr, Ni and Ca, respectively. The higher concentrations were obtained for Mn, followed by Cr, then Fe. By comparing the obtained contents with MRL (Maximum Residual Limit) for some metals (Fe, Zn, Mn, Co, Cr), the obtained results of heavy metals in this study were low comparing with (MRL) , except for manganese contents, where the results of this study recorded contamination and toxicity the studied samples by Mn, where its values (3.41 ± 1.87 µg/g) are high than MRL values, (Table2).

Table 2. The contents of Fe, Zn, Mn, and Al in tea samples.

Metal Sample No	Fe	Zn	Mn	Al
1	0.41	0.69	1.74	2.35
2	0.36	0.55	1.68	2.02
3	0.88	0.47	4.03	4.14
4	1.08	0.79	3.96	2.59
5	0.41	2.85	1.75	2.19
6	0.37	0.42	2.09	3.03
7	0.71	0.34	6.48	2.96
8	0.48	0.40	6.69	6.02
9	0.27	0.23	1.83	2.31
10	0.64	1.44	5.26	3.52
11	0.24	0.22	2.10	2.34
12	0.72	0.29	3.35	1.80
±SD	0.259	0.748	1.87	1.17
Average	0.54	0.72 1	3.41	2.93

**Figure 1. The contents of (Fe, Zn, Mn, and Al) of the studied tea samples****Table 3. The contents of Cu, Cr, Ni and Ca in the studied tea samples.**

Metal Sample No	Cu	Cr	Ni	Ca
1	0.11	0.011	ND	25.80
2	0.12	0.010	ND	21.83
3	0.08	0.012	0.015	26.11
4	0.062	0.013	0.014	19.16
5	0.10	0.014	ND	27.87
6	0.13	0.012	0.016	22.83
7	0.07	0.023	0.022	22.05
8	0.060	0.013	0.020	21.43
9	0.061	0.014	ND	17.93
10	0.10	0.021	0.013	20.6
11	0.063	0.011	ND	16.97
12	0.060	0.012	0.011	26.58
±SD	0.025	4.037	0.008	3.53
Average	0.084	1.17	0.009	22.43

ND: Non detected

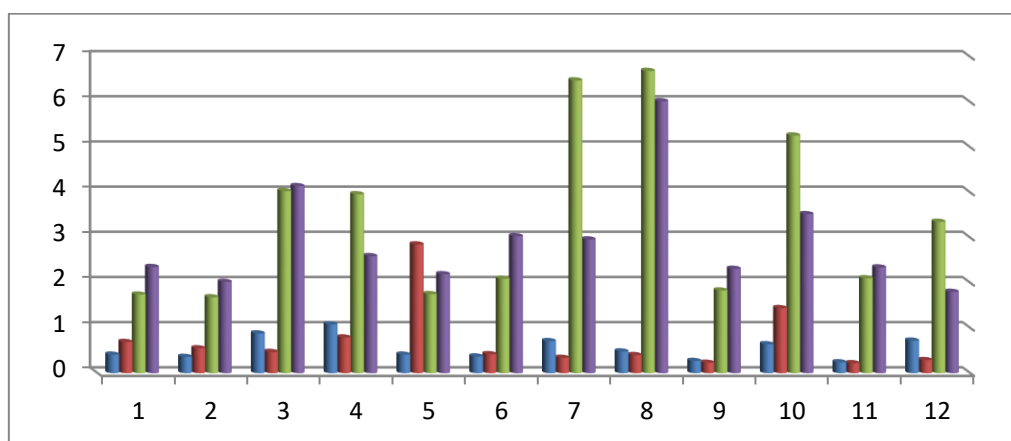


Figure 2. The contents of (Cu, Ni, and Cr) of the studied tea samples

Table 3. Compared the detected heavy metals with MRL values

Metal	Zn	Fe	Cu	Mn	Cr	Ni
Concentration	0.72 ± 0.7	0.54 ± 0.25	0.084± 0.025	3.41± 1.87	1.17± 0.4	0.009 ±0.008
MRL s	100	500	0.01	0.16	2.3	-

MRL: Minimum Residual Limit

By using diagrams (Figures 3 - 14) to evaluate higher contents of detected heavy metals in the studied samples, the results recorded all samples containing high concentrations of Mn, then the other metals were varied from one sample to another. Also, the results recorded variations of the percentage distribution of metals between the studied, the major metals in tea samples are Aluminum (Al) and manganese (Mn), besides the presence of some relatively high amounts of Zn and Fe. The Tea samples numbers of (1, 2, 3, 6, 9 and 11) showed higher contents of Al percentage values of (44, 43, 43, 50, 49 and 47 %), also there are high percentage values of Mn (33, 35, 42, 35, 39 and 42 %) from the total amount of detected heavy metals in tea samples of this study. On the side, the higher contents of Mn were recorded in the samples of (4, 5, 7, 8, 10, and 12) with values of (47, 60, 49, 48, and 54 %). Small percentage values of the other metals, Zn, Cu, Ni and Cr were observed. Relative increase in Fe contents as was recorded in tea samples.

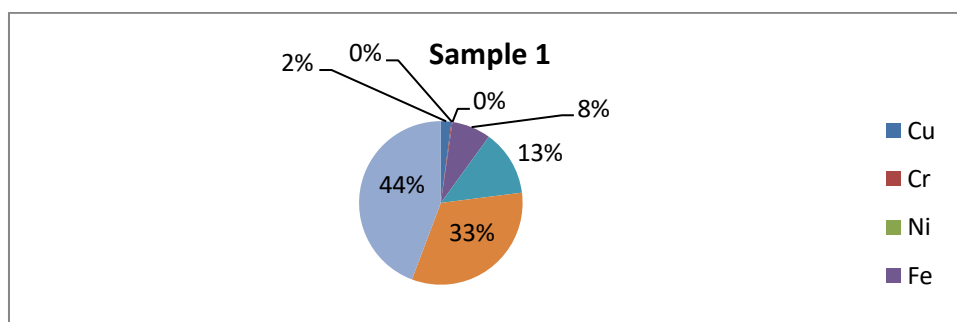


Figure 3. Pia gram of metal distribution of sample 1

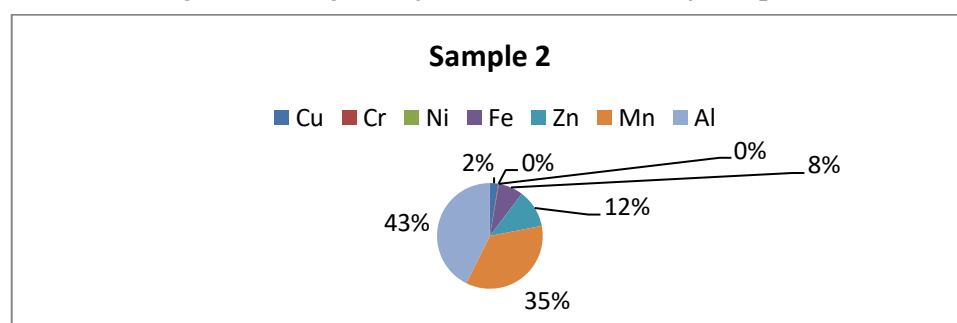
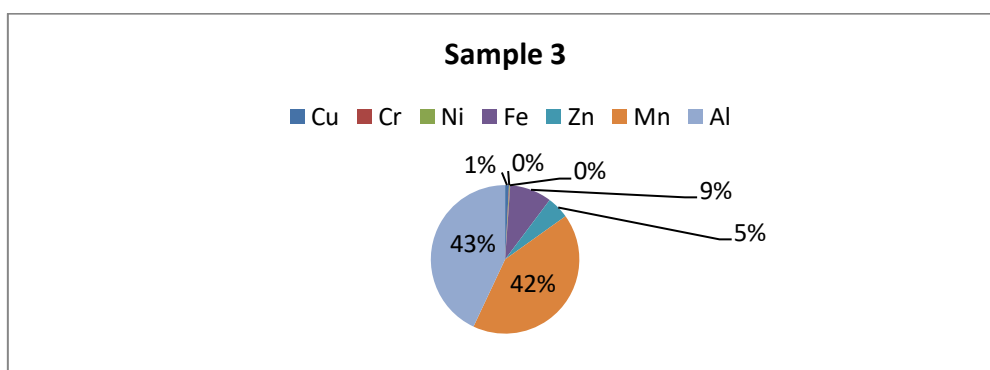
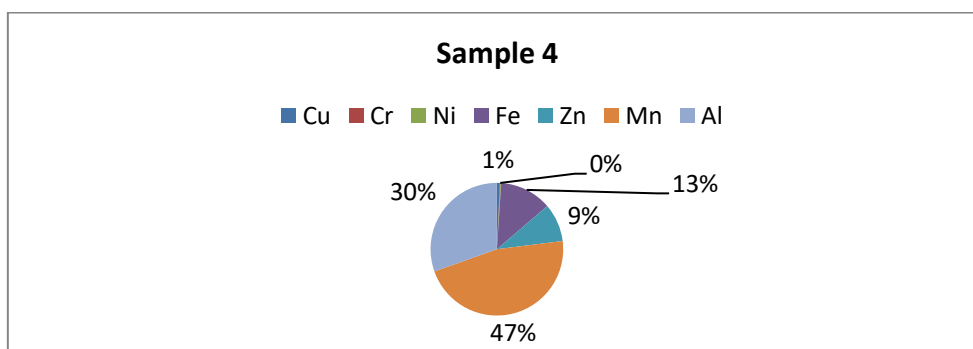
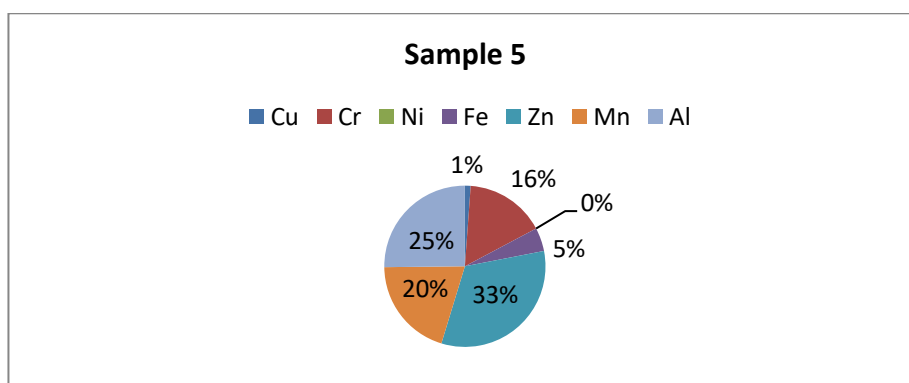
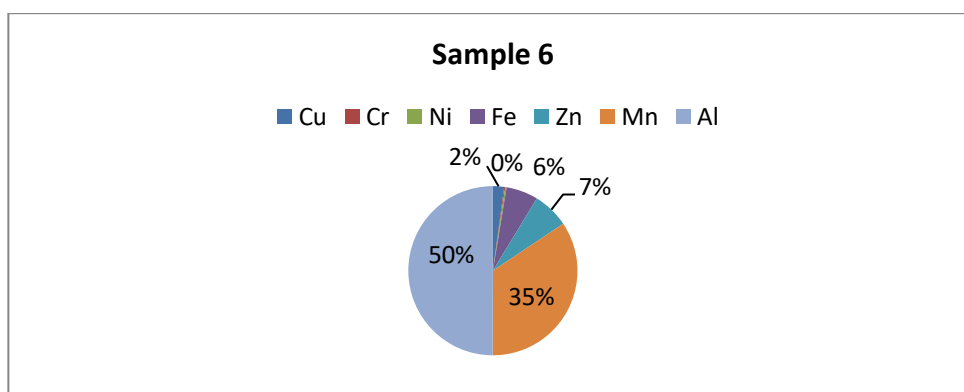
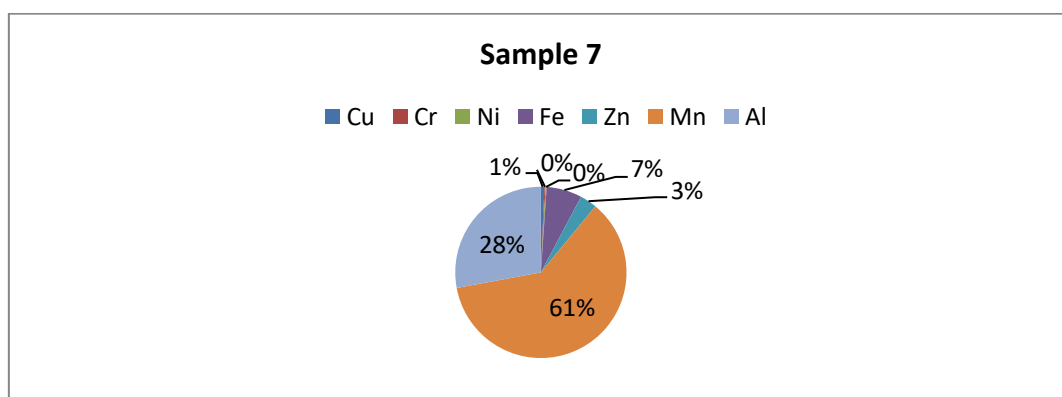
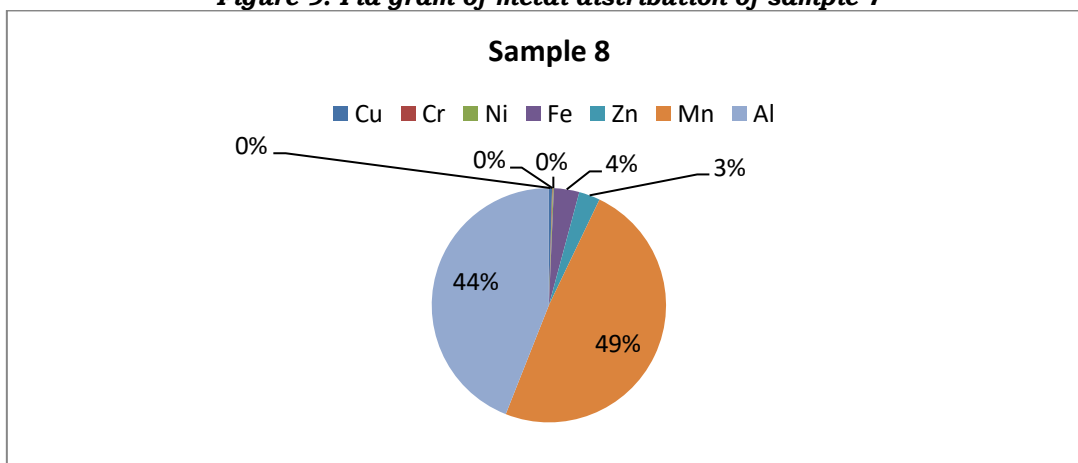
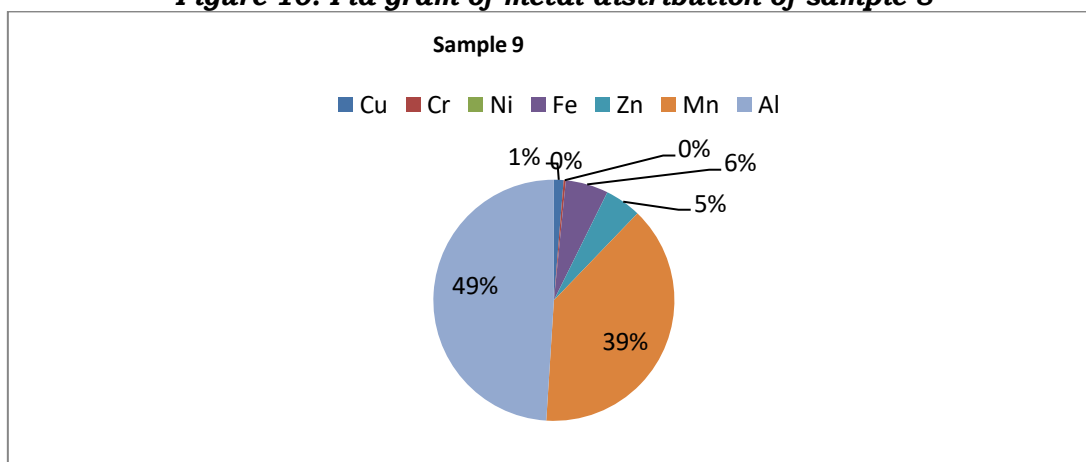
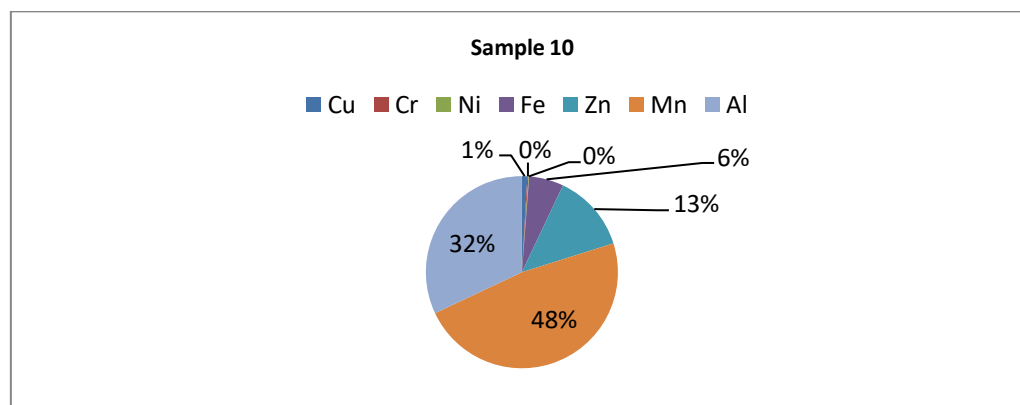


Figure 4. Pia gram of metal distribution of sample 2

**Figure 5. Pia gram of metal distribution of sample 3****Figure 6. Pia gram of metal distribution of sample 4****Figure 7. Pia gram of metal distribution of sample 5****Figure 8. Pia gram of metal distribution of sample 6**

**Figure 9. Pia gram of metal distribution of sample 7****Figure 10. Pia gram of metal distribution of sample 8****Figure 11. Pia gram of metal distribution of sample 9****Figure 12. Pia gram of metal distribution of sample 10**

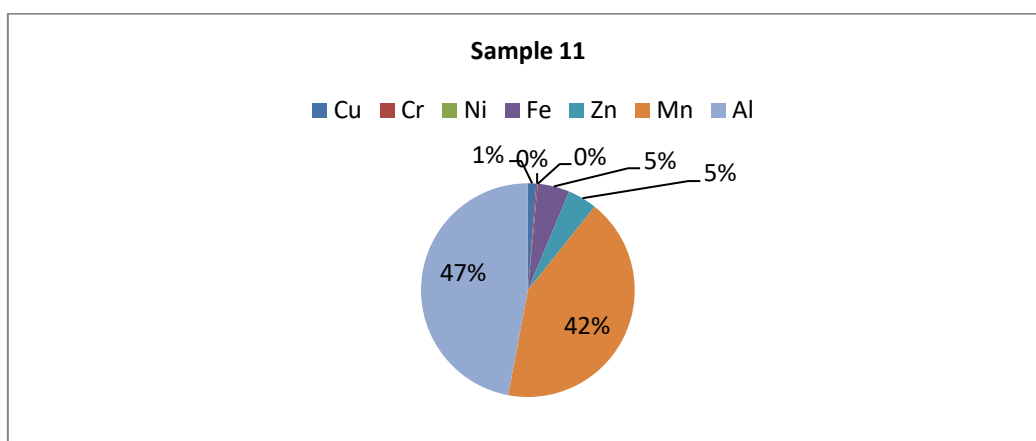


Figure 13. Pia gram of metal distribution of sample 11

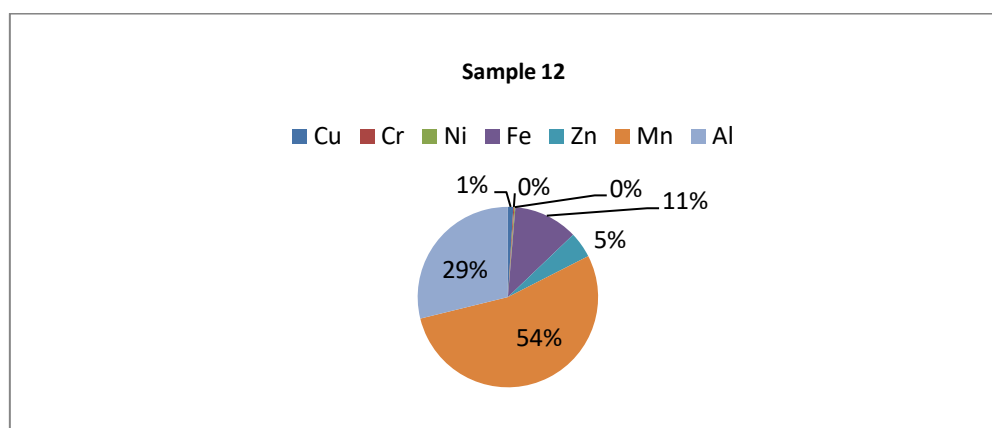


Figure 14. Pia gram of metal distribution of sample 12

Discussion

This study aimed to detect and quantify the levels of heavy metals in a diverse range of tea samples. These samples were collected through a rigorous random sampling process, encompassing both locally sourced tea and branded products. By analyzing these samples, the study sought to provide valuable insights into the potential exposure to heavy metals through tea consumption and to inform strategies for ensuring the safety and quality of this popular beverage. The analyzed tea samples exhibited varying levels of heavy metal accumulation. For instance, numerous studies have documented the presence of copper in tea samples originating from diverse geographical locations, including Iran, Lithuania, and China [95], potassium in Pakistani tea, and lead in Tunisian tea [96]. These findings underscore the metal-accumulating potential of tea plants. Additionally, other studies have reported the accumulation of aluminum and iron in tea leaves.

These observations highlight the potential for heavy metal contamination in tea, emphasizing the importance of stringent quality control measures to safeguard consumer health and well-being. The concentration of Al, Mn, and Fe in the Libyan tea samples was notably higher compared to other metals. This elevation may be attributed to the elevated metal content of the soil in the tea-producing regions and potentially to selective absorption by the tea plant. The low solubility of these metals suggests they are bound in less soluble complexes. The variability in metal content across different tea brands can be linked to their geographical origin, influenced by soil composition and leaching characteristics. Long-term tea cultivation can lead to soil acidification and increased levels of bioavailable heavy metals, potentially contributing to higher heavy metal accumulation in tea leaves [97]. The observed variations in the current study may be attributed to differences in the agro-climatic conditions of the imported tea. While the toxic effects of heavy metals have been extensively documented, the therapeutic efficacy of tea is contingent upon the concentration of its bioactive compounds.

It is imperative to maintain a delicate balance between a concentration that is sufficiently high to elicit beneficial effects and one that is low enough to avoid potential toxicity. A narrow therapeutic window exists within which the desired therapeutic outcomes can be achieved without compromising safety. The aqueous tea extracts contain appreciable quantities of metal ions that can potentially contribute to daily dietary intake. However, these levels remain below the established daily dietary requirements for humans. The presence of metal ions in tea extracts is a natural consequence of the plant's interaction with the soil and water environment [98]. These ions, such as Fe, K, Mn, and Ca, are essential micronutrients that play crucial roles in various physiological processes. While the concentrations of these ions in tea extracts may

vary depending on the type of tea, growing conditions, and processing methods, they generally fall within safe limits for human consumption. The accurate quantification of these elements in beverages, water, food, plant, and soil samples is therefore a crucial task. Green leafy vegetables represent a significant dietary source of these metals.

To ensure the safety and efficacy of tea consumption, it is recommended to regularly consume aqueous extracts for their essential nutrients. However, it is crucial to monitor and analyze tea samples from Libya and other regions to prevent exceeding daily intake limits of potentially harmful substances. Geographical variations in heavy metal concentrations have been observed among different tea brands, highlighting the importance of regular testing and quality control measures [99]. The observed inconsistencies in metal concentrations across diverse growing regions can be attributed to the inherent differences in the underlying chemical and physical properties of these areas. These variations may include soil type, mineral content, water quality, climate, and agricultural practices, all of which can significantly influence the accumulation of metals in plant tissues. Routinely consuming aqueous extracts for essential nutrients is also recommended. Regular monitoring and frequent analysis of tea in Nigeria and other regions are necessary to prevent exceeding daily intake limits.

In conclusion, the geographical origin significantly influenced the heavy metal concentrations across different tea brands. Notably, we observed substantial differences in the water solubility of these metals. Previous research has reported similar variations in metal concentrations among different tea brands. The observed wide range of metal concentrations could be attributed to the distinct chemical and physical properties of the growing regions [98-99].

Health risk assessments

To assess potential health risks associated with heavy metal exposure, THQ and HI were calculated. A THQ or HI value exceeding 1 indicates a potential health risk. Cobalt was detected in one sample, collected from a source containing Zn, Mn, and Cr at concentrations of 5.19, 1.27, and 1.19 ppm, respectively. The calculated THQ for this sample exceeded 1, suggesting a potential health risk associated with cobalt consumption. Chromium was detected in tea samples with a mean concentration of 1.19 ppm, which falls within acceptable limits. The THQ limit was exceeded in our study [1], levels of cobalt and manganese, both of which are known to be highly toxic and may pose significant long-term health risks.

Table 4. The results of the THQ values

Metal	THQ
Cu	0.138
Cr	1.19
Ni	0.014
Ca	37.003
Fe	0.89
Zn	1.27
Mn	5.18
Al	4.83

Mn levels exceeded MRLs in several samples, with different concentrations of Zn, Fe, Al, Cu, Cr, Ni, and Ca within permissible limits. THQ values for Mn and Cr surpassed 1 in some samples, indicating potential non-carcinogenic risks. HI values also exceeded 1 in select samples, suggesting cumulative health concerns.

Conclusion

The heavy metals recorded in this study include different types of heavy metals such as Fe, Zn, Cr, Mn, Ni, and Cu, besides the presence of different concentrations of Al and Ca. The risk indexes showed an increase in the values of Cr, Zn, and Mn compared with safety limits.

Acknowledgment

The authors highly appreciated the collaboration of the central laboratory of chemical analysis at Omar Al-Mukhtar University during sample preparation for the establishment of this study.

Conflict

No conflict of the results of this study with other studies.

References

1. Lambdacyhalothrin (General Fact Sheet). Corvallis (OR): National Pesticide Information Center (NPIC). Available from: <https://npic.orst.edu/factsheets/lambda.html>

2. Chen Z. Pesticide residue in tea and its risk assessment. Hangzhou: Tea Research Institute, Chinese Academy of Agricultural Sciences; 2013.
3. Soomro MT, Zahir E, Mohiuddin S, Khan N, Naqvi II. Quantitative assessment of metals in local brands of tea in Pakistan. *Pak J Biol Sci.* 2007;10(2):1-5.
4. Ebadi AG, Zare S, Mahdavi M, Babaee M. Study and measurement of Pb, Cd, Cr and Zn in green leaf of tea cultivated in Gillan province of Iran. *Pak J Nutr.* 2005;4(4):270-2.
5. Atta MB. Aluminum content in dust black tea leaves and its beverages. *Menofiya J Agric Res.* 1995;20(4):137-50.
6. Lee HC. Review of inductively coupled plasmas: Nano-applications and bistable hysteresis physics. *Appl Phys Rev.* 2018;5(1):011108. DOI: 10.1063/1.5012001.
7. Trejos T, Corzo R, Subedi K, Almirall JR. Elemental analysis of glass and paint materials by Laser Ablation Inductively Coupled Plasma Mass Spectrometry (LA-ICP-MS) for forensic application. 2006.
8. Hamad IH, Nuesry MS. The poly cyclic hydrocarbons levels in some fishes tissues collected from Derna City (Libya) Coast. In: International conference on chemical, agricultural and medical sciences; 2014 Dec 4-5; Antalya, Turkey. 2014. p. 52-6.
9. Hamad MAH, Mounera AAE, Baseet ESM, Eman E, Al-Badri M. Identification and detection the aromatic and aliphatic hydrocarbons in Epinephelus Marginatus fish samples collected from Benghazi coast. *Int J Adv Multidiscip Res Stud.* 2023;6(3):107-13.
10. Mohammed A, Hamad MAH, Mounera AAE, Eman IHE. Extraction and identification of aliphatic hydrocarbons in marine sediment samples at Benghazi city and Dyriana town coasts (Libya). *J Res Humanit Soc Sci.* 2023;11(10):168-74.
11. Hasan MAH, Muftah HS, Abdelghani KA, Saad SI. Poly aromatic hydrocarbon concentrations in some shell samples at some Tobrouk city coast regions: could the oil industry be significantly affecting the environment. *Ukr J Ecol.* 2022;12(3):21-8.
12. Habel AMA, Mohamed NIH, Mohammed MA, Hamad MAH. The levels and sources of aliphatic and polycyclic aromatic hydrocarbons in blue runner fish from Benghazi coast, Libya. *Afr J Biol Sci.* 2024;6(3):1-10.
13. Hasan HMI, Mohamad ASA. A study of aliphatic hydrocarbons levels of some waters and sediments at Al-Gabal Al-Akhder coast regions. *Int J Chem Sci.* 2013;11(2):833-49.
14. Salem GM, Aljidaemi FF, Hwisa SA, Hamad MIH, Zaid AA, Amer IO. Occupational exposure to benzene and changes in hematological parameters in East Tripoli, Libya. *Nanotechnol Percept.* 2024;20(S5):358-64.
15. Habil Z, Ben arous N, Masoud N, Hasan H. Using GC-mass method for determination the hydrocarbon compounds in some vegetable samples at Derna city, Libya. *Libyan Med J.* 2025;17(3):374-83.
16. Hamad R, Ikraiam F, Hasan H. Determination of specific natural radionuclides in the bones of some local fish commonly consumed from the eastern Libyan coast. *J Rad Nucl Appl.* 2023;8(3):283-9.
17. Sroor AT, Walley El-Dine N, El-Bahi SM, Hasa HMA, Ali JM. Determination of radionuclides levels and absorbed dose for soil, rock, plant and water in gondola- Libya. *IOSR J Appl Phys.* 2018;10(4):40-9.
18. Hayder AMS, Hasan HM, Ikraiam F. Determination and comparative analysis of natural radioactivity levels using gamma spectrometry in shore sediment samples from the east coast, Libya. In: Sebha University conference proceedings; 2024; Sebha, Libya. 2024. p. 297-300.
19. Hasan H, Ammhamid R, Khatab H, Ali J, Al kaseh A. Using gamma ray radiation to estimate the types and contents of radioactive nuclides in some ported sugar samples, Libya. *AlQalam J Med Appl Sci.* 2025;8(3):1795-803.
20. Eltawaty SA, Abdalkader GA, Hasan HM, Houssein MA. Antibacterial activity and GC-MS analysis of chloroform extract of bark of the Libyan *Salvia fruticosa* Mill. *Int J Multidiscip Sci Adv Technol.* 2021;1(1):715-21.
21. Aljamal MA, Hasan HM, Al Sonosy HA. Antibacterial activity investigation and anti-biotic sensitive's for different solvents (Ethanol, propanol, DMSO and di Ethel ether) extracts of seeds, leafs and stems of (*Laurus azorica* and *Avena sterilis*) plants. *Int J Curr Microbiol App Sci.* 2024;13(11):175-90.
22. Hamade MH, Abdelraziq SA, Gebreel AA. Extraction and determination the of Beta carotene content in carrots and tomato samples collected from some markets at ElBeida City, Libya. *EPH Int J Appl Sci.* 2019;1(1):105-10.
23. Hasan HM, Ibrahim H, Gonaïd MA, Mojahidul I. Comparative phytochemical and antimicrobial investigation of some plants growing in Al Jabal Al-Akhder. *J Nat Prod Plant Resour.* 2011;1(1):15-23.
24. Hasan H, Jadallah S, Zuhir A, Ali F, Saber M. The Anti-Cancer, Anti-Inflammatory, Antibacterial, Antifungal, Anti-Oxidant and phytochemical investigation of flowers and stems of *Anacyclus Clavatus* plant extracts. *AlQalam J Med Appl Sci.* 2025;8(3):415-27.
25. Hasan H, Zuhir A, Shuib F, Abdraba D. Phytochemical investigation and exploring the *Citrullus Colocynthis* extracts as antibacterial agents against some gram and negative bacteria species. *AlQalam J Med Appl Sci.* 2025;8(3):392-400.
26. MdZeyauallah R, Naseem A, Badrul I, Hamad MI, Azza SA, Faheem AB, et al. Catechol biodegradation by *Pseudomonas* strain: a critical analysis. *Int J Chem Sci.* 2009;7(3):2211-21.
27. El-Mehdawy MF, Eman KS, Hamad MI, Hasan H. Amino acids contents of leafs and stems for two types of herbal plants (Marjoram and Hybrid tea rose) at AL-Gabal AL-Akhder region. *Der Pharma Chem.* 2014;6(6):442-7.
28. El-Mehdawy MF, Eman KS, Hamad MIH. Amino acid contents of leafs and stems for three types of herbal plants at AL-Gabal AL-Akhder region. *World J Chem.* 2014;9(1):15-9.
29. Hamad MH, Noura AAM, Salem AM. Phytochemical screening, total phenolic, anti-oxidant, metal and mineral contents in some parts of plantago Albicans grown in Libya. *World J Pharm Res.* 2024;13(3):1-17.
30. Anees AS, Hamad MIH, Hasan H, Mojahidul I. Antifungal potential of 1,2-4triazole derivatives and therapeutic efficacy of *Tinea corporis* in albino rats. *Der Pharm Lett.* 2011;3(1):228-36.

31. Hamad Hasan, Marwa Mohammed, Amal Haroon. Determining the contents of antioxidants, total phenols, carbohydrate, total protein, and some elements in *Eucalyptus gomphocephala* and *Ricinus communis* plant samples. *Libyan Med J.* 2015;1(1):222-31.
32. Hamad Hasan, Zuhir Akrim, Farag Shuib, Dala Abdraba. Efficiency of *Cynara Cornigera* fruits on antibacterial, antifungal and its phytochemical, anti-oxidant screening. *Libyan Med J.* 2025;3(1):120-8.
33. Hamad Hasan, Ashour Sulayman, Ahmed Alehrir. Estimation of amino acid composition, total carbohydrate, and total protein content in *Ballota pseudodictamnus* plant extracts from Al Jabal Al Akhdar Region, Libya. *Libyan Med J.* 2025;3(1):266-71.
34. Hamad Hasan, Ahmed Hamad, Wafa Abdelsatar. Evaluation of anti-oxidant capacity, total phenol, metal, and mineral contents of *Ziziphus lotus* plant grown at some regions of AlGabal AlKhder, Libya. *Libyan Med J.* 2025;3(1):137-43.
35. Hesien RA, Amira AKA, Ahlaam MA, Hamad MAH. Determination the anti-oxidant capacity, total phenols, minerals and evaluation the anti- bacteria activity of leafs and stems of *Gaper* plant extracts. *Sch J Appl Med Sci.* 2024;12(4):451-7.
36. Ben Arous NA, Naser ME, Hamad MAH. Phytochemical screening, anti-bacterial and anti-fungi activities of leafs, stems and roots of *C. parviflorus* Lam and *C. salviifolius* L plants. *Int J Curr Microbiol App Sci.* 2014;13(11):262-80.
37. Anas FAE, Hamad MAH, Salim AM, Azza MH. Phytochemical screening, total phenolics, antioxidant activity and minerals composition of *Helichrysum stoechas* grown in Libya. *Afr J Biol Sci.* 2024;3(6):2349-60.
38. Naseer RE, Najat MAB, Salma AA, Hamad MAH. Evaluation of metal and mineral contents of leafs, stems and roots of *C. Parviflorus* Lam and *C. Salviifolius* L plants growing at Al Ghabal Al-Khder (Libya). *Int J Adv Multidiscip Res Stud.* 2024;4(5):191-4.
39. Hamad MAH, Salem AM. Total carbohydrate, total protein, minerals and amino acid contents in fruits, pulps and seeds of some cultivars of muskmelon and watermelon fruit samples collected from Algabal Alkhder region. *Sch J Appl Med Sci.* 2024;12(1):1-7.
40. Gonaid MI, Ibrahim H, Al-Arefy HM. Comparative chemical and biological studies of *Salvia fruticosa*, *Ocimum basillicum* and *Pelargonium graveolans* cultivated in Al-Jabal Al- AkhdarAkhdar. *J Nat Prod Plant Resour.* 2012;6(2):705-10.
41. Rinya FMA, Hamad MAH, Ahlam KA, Hammida MEH. Phytochemical screening of some herbal plants (*Menthe*, *Origanum* and *Salvia*) growing at Al-Gabal Al-akhder Region-Libya. *Afr J Basic Appl Sci.* 2017;9(3):161-4.
42. Anas FAA, Hamad MAH, Salim AA, Azza MH. Phytochemical screening, total phenolics, antioxidant activity and minerals composition of *Helichrysum stoechas* grown in Libya. *Afr J Biol Sci.* 2024;3(6):2349-60.
43. Haroon A, Hasan H, Wafa AAS, Baset ESM. A comparative study of morphological, physiological and chemical properties of leafs and steam samples of (*E.gomphocephala*) (Tuart) plant growing at coastal (Derna city) and *J Res Environ Earth Sci.* 2024;9(12):10-8.
44. Hamad MAS, Ali AR. Separation and identification the speciation of the phenolic compounds in fruits and leaves of some medicinal plants (*Juniperus phoenicea* and *Quercus coccifera*) growing at Algabal Al Akhder region, Libya. *Indian J Pharm Educ Res.* 2016;51(3):299-303.
45. Enam FM, Wesam FAM, Hamad MAH. Detection the contents of minerals of (Sodium, Potassium and Calcium) and some metals of (Iron, Nickel and Copper) in some vegetable and soil samples collected from Al-Marj. *Int J Adv Multidiscip Res Stud.* 2023;5(3):304-9.
46. Hamad MIH, Safa RM Mousa. Synthesis and (IR and TEM) characterization of leafs and stem nanoparticles of *Artemisia* plant: comparative study for the evaluation of anti-bacterial efficiency. *Int J Adv Multidiscip Res Stud.* 2024;4(5):195-9.
47. Elsalhin H, Abobaker HA, Hasan H, El-Dayek GA. Antioxidant capacity and total phenolic compounds of some algae species (*Anabaena* and *Spirulina platensis*). *Sch Acad J Biosci.* 2016;4(10):782-6.
48. Alaila AK, El Salhin HE, Ali RF, Hasan HM. Phytochemical screening of some herbal plants (*Menthe*, *Origanum* and *Salvia*) growing at al-gabal al-akhder region- Libya. *Int J Pharm Life Sci.* 2017;8(4):5500-3.
49. Hasan H, Mariea FFE, Eman KS. The contents of some chemical compounds of leafs and stems of some herbal plants (*Thymy*, *Rosemary*, *Salvia*, *Marjoram* and *Hybrid Tea Rose*) at Al-Gabal Al-Akhder region. *EPH Int J Appl Sci.* 2014;6(3):1-8.
50. Abdelrazeg A, Khalifa A, Mohammed H, Miftah H, Hamad H. Using melon and watermelon peels for the removal of some heavy metals from aqueous solutions. *AlQalam J Med Appl Sci.* 2025;8(3):787-96.
51. Abdul Razaq A, Hamad H. Estimate the contents and types of water well salts by the Palmer Roger model affecting the corrosion of Al-Bayda city (Libya) network pipes. *AlQalam J Med Appl Sci.* 2025;8(3):744-53.
52. Abdulsayid FA, Hamad MAH, Huda AE. IR spectroscopic investigation, X-ray fluorescence scanning, and flame photometer analysis for sediments and rock samples of Al-Gabal Al-Akhder coast region (Libya). *IOSR J Appl Chem.* 2021;14(4):20-30.
53. ALambarki M, Hasan HMA. Assessment of the heavy metal contents in air samples collected from the area extended between Albayda and Alquba cities (Libya). *AlQalam J Med Appl Sci.* 2025;8(3):695-707.
54. Al-Nayyan N, Mohammed B, Hamad H. Estimate of the concentrations of heavy metals in soil and some plant samples collected from (near and far away) of the main road between Al-Bayda city and Wadi Al-Kouf region. *AlQalam J Med Appl Sci.* 2025;8(3):816-26.
55. Hasan HMI. Studies on physicochemical parameters and water treatment for some localities along coast of Alexandria [Doctoral dissertation]. [Alexandria, Egypt]: Alexandria University; 2006.
56. Hamad MAH, Hager AA, Mohammed EY. Chemical studies of water samples collected from area extended between Ras Al-Halal and El Hanica, Libya. *Asian J Appl Chem Res.* 2022;12(3):33-46.
57. Hamad M, Mohammed AA, Hamad MAH. Adsorption and kinetic study for removal some heavy metals by use in activated carbon of sea grasses. *Int J Adv Multidiscip Res Stud.* 2024;4(6):677-85.

58. Hamad MAH, Hamad NI, Mohammed MYA, Hajir OAA, Al-Hen dawi RA. Using bottom marine sediments as environmental indicator state of (Tolmaitha – Toukra) region at eastern north coast of Libya. *Sch J Eng Tech.* 2024;2(14):118-32.
59. Hamad MIH. The heavy metals distribution at coastal water of Derna city (Libya). *Egypt J Aquat Res.* 2008;34(4):35-52.
60. Hamad MIH, Mojahid ul Islam. The concentrations of some heavy metals of Al-Gabal Al-Akhdar Coast Sediment. *Arch Appl Sci Res.* 2010;2(6):59-67.
61. Hamad MAH, Amira AKA. Estimate the concentrations of some heavy metals in some shoes polish samples. *EPH Int J Appl Sci.* 2016;2(2):24-7.
62. Hamad MAH, Hussien SSM, Basit EEM. Accumulation of some heavy metals in green algae as bio indicators of environmental pollution at Al-Hania region: Libya coastline. *Int J Adv Multidiscip Res Stud.* 2024;4(5):188-90.
63. Hamad MIH, Ahmed MA. Major cations levels studies in surface coastal waters of Derna city, Libya. *Egypt J Aquat Res.* 2009;35(1):13-20.
64. Hamad MIH, Masoud MS. Thermal analysis (TGA), diffraction thermal analysis (DTA), infrared and X-rays analysis for sediment samples of Toubrouk city (Libya) coast. *Int J Chem Sci.* 2014;12(1):11-22.
65. Hamad R, Ikraiam FA, Hasan H. Estimation of heavy metals in the bones of selected commercial fish from the eastern Libyan coast. *J Rad Nucl Appl.* 2024;9(1):47-51.
66. Hasan HAH. Estimate lead and cadmium contents of some archeological samples collected from ancient cities location (Cyrene and Abolonia) at Al-Gabal Al-Akhder Region, Libya. *Univ J Chem Appl.* 2021;12(21):902-7.
67. Alfutisi H, Hasan H. Removing of thymol blue from aqueous solutions by pomegranate peel. *EPH Int J Appl Sci.* 2019;1(1):111-9.
68. Hasan JA, Hasan HMA. Potential human health risks assessment through determination of heavy metals contents in regularly consumed yogurta in Libya. *World J Pharm Pharm Sci.* 2024;13(12):100-12.
69. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Essam AM, Hamad MIH. Rice husk and activated carbon for waste water treatment of El-Mex Bay, Alexandria Coast, Egypt. *Arab J Chem.* 2016;9(S2):S1590-S6.
70. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Hamad IH. Heavy metals accumulation in sediments of Alexandria coastal areas. *Bull Fac Sci.* 2012;47(1-2):12-28.
71. Mamdouh SM, Wagdi ME, Ahmed MA, Hamad MIH. Chemical studies on Alexandria coast sediment. *Egypt Sci Mag.* 2005;2(4):93-102.
72. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Hamad MIH. Distribution of different metals in coastal waters of Alexandria, Egypt. *Egypt Sci Mag.* 2010;7(1):1-19.
73. Mohamed AE, Afnan SA, Hamad MA, Mohammed AA, Mamdouh SM, Alaa RE, et al. Usage of natural wastes from animal and plant origins as adsorbents for the removal of some toxic industrial dyes and heavy metals in aqueous media. *J Water Process Eng.* 2023;55:104192.
74. Mohamed HB, Mohammed AZ, Ahmed MD, Hamad MAH, Doaa AE. Soil heavy metal pollution and the associated toxicity risk assessment in Ajdabiya and Zueitina, Libya. *Sci J Damietta Fac Sci.* 2024;14(1):16-27.
75. Nabil B, Hamad H, Ahmed E. Determination of Cu, Co and Pb in selected frozen fish tissues collected from Benghazi markets in Libya. *Chem Methodol.* 2018;2:56-63.
76. Hamad Hasan, Aljidaemi F, Salem G, Bannur Z, Aljeaday H, Aldaha L, et al. Utilization of Chrysanthemum leaf and stem extracts as antioxidant, anticancer, anti-inflammatory, and phytochemical agents. *Asian J Green Chem.* 2026;10:84-100.
77. Wesam FAM, Hamad MAH. Study the accumulation of minerals and heavy metals in Ulva algae, Cladophora, Polysiphonia and Laurencia algae samples at eastern north region of Libya coast. *GSC Biol Pharm Sci.* 2023;23(03):147-52.
78. Citrine E, Hamad H, Hajer Af. Contents of metal oxides in marine sediment and rock samples from the eastern Libyan coast, utilizing the X-ray method. *AlQalam J Med Appl Sci.* 2015;1(1):1316-21.
79. Hanan MA, Hamida E, Hamad MAH. Nitrogen, phosphorus and minerals (Sodium, Potassium and Calcium) contents of some algae's species (Anabaena and Spirulina platensis). *Int J Curr Microbiol App Sci.* 2016;5(11):836-41.
80. Mardhiyah F, Hamad H. Assessment of soil contamination by heavy metals in the Al-Fatayeh Region, Derna, Libya. *AlQalam J Med Appl Sci.* 2025;8(3):1081-91.
81. Hamad MIH, Aaza IY, Safaa SHN, Mabrouk MS. Biological study of transition metal complexes with adenine ligand. *Proc.* 2019;41(1):77.
82. Ahmed ONH, Hamad MAH, Fatin ME. Chemical and biological study of some transition metal complexes with guanine as ligand. *Int J New Chem.* 2023;10(3):172-83.
83. Hamad MAH, Enas UE, Hanan AK, Hana FS, Somia MAE. Synthesis, characterization and antibacterial applications of compounds produced by reaction between Barbitol with Threonine, glycine, lycine, and alanine. *Afr J Biol Sci.* 2024;6(4):1-10.
84. Hasan H, Habil Z, Ben arous N. Estimate the types and contents of phenolic acid in C.Paviflorus lam and C.salviifolius L plants growing at Al –Gabal Al-hder regions. *AlQalam J Med Appl Sci.* 2025;8(3):1646-56.
85. Emrayed H, Hasan H, Liser R. Corrosion inhibition of carbon steel using (Arginine –levofloacin-metal) complexes in acidic media. *AlQalam J Med Appl Sci.* 2025;8(3):1633-40.
86. Hasan H, Abdelgader I, Emrayed H, Abdel-Gany K. Removal of the medical dye safranin from aqueous solutions by sea grasses activated carbon: a kinetic study. *AlQalam J Med Appl Sci.* 2025;8(3):428-34.
87. Hasan HMA, Alhamdy MA. Adsorption and kinetic study for removal some heavy metals by using activated carbon of sea grasses. *Int J Adv Multidiscip Res Stud.* 2024;4(6):677-85.
88. Almadani EA, Hamad MAH, Kwakab FS. Kinetic study of the adsorption of the removal of bromo cresol purple from aqueous solutions. *Int J Res Granthaalayah.* 2019;7(12):1-10.

89. Mamdouh SM, Wagdi ME, Ahmed MA, Alaa EA, Essam AM, Hamad MIH. Rice husk and activated carbon for waste water treatment of El-Mex Bay, Alexandria Coast, Egypt. Arab J Chem. 2016;9(S2):S1590-S6.
90. Hasan H, Ammhmmd R, Khatab H, Ali J, Al kaseh A. Using gamma ray radiation to estimate the types and contents of radioactive nuclides in some ported sugar samples, Libya. AlQalam J Med Appl Sci. 2025;8(3):1795-803.
91. Abdull-Jalliel H, B Arous N, Alhoreir M, Hasan H. Using the extracts of the (Dodder) plant and the concentrations of some metals as inhibitors for growth, the (Pseudomonas) bacteria isolated from some hospital rooms in Derna and Al bayda. AlQalam J Med Appl Sci. 2025;8(3):1600-11.
92. Balal A, Obid M, Khatab H, Hasan H. Determination of lead and cadmium marine water and crabs (pachygrapsus marmoratus) from tolmitha coast, Libya. AlQalam J Med Appl Sci. 2025;8(3):1670-7.
93. Abdull-Jalliel H, Sulayman A, Alhoreir M, Hasan H. The antimicrobial effect of some metal concentration on growth of staphylococcus and klebsiella bacteria species. AlQalam J Med Appl Sci. 2025;8(3):1646-56.
94. Ansari F, Norbaksh R, Daneshmandirani K. Determination of heavy metals in Iranian and imported black tea. Iran J Environ Health Sci Eng. 2007;4(4):243-8.
95. Tautkus S, Kazlauskas R, Kareiva A. Determination of copper in tea leaves by flame atomic absorption spectrometry. CHEMIJA. 2004;15(4):49-52.
96. Hamdaoui M, Chahed A, Ellouze-Chabchoud S, Marouani N, Abid ZB, Hadhili A. Effect of green tea decoction on long-term iron, zinc and selenium status of rats. Ann Nutr Metab. 2005;49(2):118-24.
97. Zhang M, Zhou C, Huang C. Relationship between extractable metals in acid soils and metals taken up by tea plants. 2006 [cited year unknown]. Available from: http://cat.inist.fr/?a_mode=afficheN&cpsidt=17561212
98. Gramza A, Wojciak RW, Korczak J, Hes M, Wisniewska J, Krejpcio Z. Influence of the Fe and Cu presence in tea extracts on antioxidant activity. Electron J Pol Agric Univ. 2005;8(4):30. Available from: <http://www.ejpau.media/pl/volume8/issue4/art-30.html>